

Inflammation in acute coronary syndromes

MARK ROBBINS, MD, AND ERIC J. TOPOL, MD

he late Russell Ross's assertion that atherosclerosis is an inflammatory disease is now strongly supported by clinical, basic, and pathological research calling for an evolution in thought concerning the evaluation and treatment of acute coronary syndromes (ACS).¹⁻⁵ The initial insult is endothelial injury and subsequent dysfunction via the deleterious effects of the known cardiac risk factors such as oxidized LDL, infection, hyperglycemia, hypertension, hyperhomocysteinemia, or smoking. Irrespective of the cause of endothelial damage, the resultant activation and proliferation of inflammatory cells, smooth muscle cells, generation of cytokines, growth factors, and many other substances lead to the progression of atherosclerosis. The presence and degree of inflammation and procoagulant state, defined by elevated CRP, fibrinogen, interleukin (IL)-1, IL-6, TNF- α , adhesion molecules, plasminogen activator inhibitor (PAI-1), tissue factor, and composition of the atherosclerotic plaque have been strongly associated with an increased risk of future cardiac events.⁶⁻⁹ Thus, the perpetuation of the inflammatory response likely plays a pivotal role in the pathobiology and vulnerability of the atherosclerotic plaque.

PATHOBIOLOGY OF INFLAMMATION, ATHEROSCLEROSIS, AND ACS Endothelial function

The endothelium lies in a critical location between the remaining vascular wall and the circulating blood thereby functioning as the pivotal barrier that protects the arterial wall from injury. This critical monolayer of cells is pluripotential, carrying out the following functions: 1) provision of a nonthrombotic surface; 2) maintenance of vascular tone through the production and release of nitric oxide (NO), prostacyclin, and endothelin; 3) regulation of growth factors and cytokines; 4) provision of a nonadherent surface for leukocytes and platelets; and 5) the modification of lipoproteins as they transverse its permeable barrier.⁵ Injury to this monolayer plays a key role in

the initiation and progression of the atherosclerotic lesion by increasing adhesive cell surface glycoproteins such as vascular cell adhesion molecule-l (VCAM-1) and intracellular adhesion molecule (ICAM), adherence, migration, and activation of leukocytes, and smooth muscle cells, production of cytokines, chemokines, and growth factors, as well as the reversal from an antithrombotic to a prothrombotic state.^{4,10-12}

Adhesion molecules

Cell-cell interactions are a vital component in the pathogenesis of inflammation. Collectively known as cell adhesion molecules, three distinct families exist—the selectins, the integrins, and the immunoglobulin superfamily each with its own specific role in the inflammatory process. The process entails tethering and rolling of leukocytes on the activated endothelium, leukocyte activation, and ultimately firm adhesion and transendothelial migration along a chemotactic gradient generated by mediators of inflammation.^{13,14}

Selectins are expressed on the cell surface of leukocvtes (L-selectin), platelets (P-selectin), and endothelial cells (E-selectin). Upon activation from inflammatory cytokines, mainly TNF- α and IL-1, cell surface expression of each selectin is enhanced.¹⁵⁻¹⁷ This process is vital in the early phase of inflammation mediating leukocyte recruitment and transient endothelial cell to leukocyte interactions (tethering and rolling phase). The subsequent steps of firm adhesion and migration of leukocytes is predominantly mediated through the interaction of integrins [leukocyte function associated antigen-1 (LAF-1), macrophage antigen-1 (MAC-1), very late activation antigen-4 (VLA-4) and GPIIb/IIIa receptor], the immunoglobulin superfamily [vascular cell adhesion molecule-1 (VCAM-1), intercellular adhesion molecule-1 (ICAM-1) and intercellular adhesion molecule-2 (ICAM-2)] and potent stimulation by inflammatory cytokines including IL-1, IL-4, IL-8, TNF- α , INF- γ , and chemokines such as chemotactic protein-1.13,14 In addition to the cell adhesion molecules on endothelial cells, leukocytes, and platelets, ICAM-1 and VCAM-1 are expressed on smooth muscle cells.¹⁸ The interaction between leukocytes and smooth muscle cells contributes to smooth muscle cell migration and proliferation, cellular composition of the atherosclerotic plaque, and an increased expression of monocyte tissue factor mRNA, all of which are likely to be vital in influencing plaque stability.^{18,19} An additional component that ties inflammation

SII-130 CLEVELAND CLINIC JOURNAL OF MEDICINE

From the Department of Cardiovascular Medicine, The Cleveland Clinic Foundation. Address correspondence to E.J.T., The Cleveland Clinic Foundation, 9500 Euclid Ave., Cleveland, Ohio 44195. E-mail: topole@ccf.org

Reprinted from Acute Coronary Syndromes, 2nd Edition, Eric J. Topol, editor, with permission of the publisher (New York: Marcel Dekker, 2001:1-31). Copyright ©2001, Marcel Dekker, Inc.

and the prothrombotic state involves the adhesion of activated platelets to the endothelium through the P-selectin-GPIIb/IIIa receptor interactions with subsequent platelet aggregation and thrombus formation.²⁰

Growing evidence supports that the presence of increased cell adhesion molecules in serum or vascular tissue may reflect ongoing active vascular remodeling due to persistent inflammation. Elevated serum levels of the soluble form of the VCAM-1 receptor (sVCAM-1) has been associated with the extent of atherosclerosis in patients with peripheral vascular disease.²¹ In patients with coronary artery disease, elevated levels of the soluble ICAM-1 (sICAM-1) has been found to be inversely proportional to HDL levels and associated with the presence of other coronary risk factors, unstable angina, myocardial infarction, and importantly to increased risk of future myocardial infarction in apparently healthy men.^{22,23} Interestingly, immunohistochemical evaluation of coronary athrectomy tissue has shown P-selectin but not E-selectin, or ICAM-1 was expressed significantly greater in the setting of unstable angina versus stable angina.²⁴ This reflects an augmented response between an endothelial cell adhesion molecule and the activated platelet linking thrombus formation and unstable coronary syndromes.

Treatment strategies available based on the inhibition of cell to cell interactions have shown promise in the treatment of chronic inflammatory diseases, and recently coronary artery disease.^{13,25} This should not be surprising given the marked similarities that exist between the pathophysiology of inflammatory diseases, such as rheumatoid arthritis, and atherosclerosis (Table 1). ASA and other NSAIDS affect the expression and function of cell adhesion molecules, and have been shown to inhibit many phases of the adhesion cascade.¹⁸ Direct antagonism via monoclonal antibodies and selectin-blocking agents against ICAM-1 and L-selectin has been shown to reduce neutrophil accumulation and myocardial injury in experimental animal studies.^{26,27} New approaches using antisense oligonucleotides to inhibit mRNA translation for cell adhesion molecule expression, and inhibition of gene expression by synthetic DNA molecules and triplex-forming oligonucleotides have shown conceptual promise in animal studies.¹³

CELLULAR AND HUMORAL MEDIATED RESPONSE Monocytes and macrophages

Monocytes, the circulating precursors of tissue macrophages, are essential in the progression of atherosclerosis and are found in all stages of atherosclerotic lesions.^{4,28} Their recruitment and infiltration through the endothelium into the intima are tightly coupled to the humoral activity of the T-lymphocyte. The colocalization of CD4+ T-cells and macrophages and the abundant expression of HLA II molecules in atherosclerotic lesions is strong evidence for the role of cell-mediated immunity in the development and progression of atherosclerosis. Population size of CD14dimCD16a+ peripheral blood monocytes has been shown to correlate with degree of hypercholesterolemia and is dramatically reduced with lipid lowering therapy.²⁹ This phenotypic expression, in con-

TABLE 1

SIMILARITIES BETWEEN ATHEROSCLEROSIS AND RHEUMATOID ARTHRITIS

	Atherosclerosis	Rheumatoid arthritis
Macrophage activation TNF-α Metalloproteinases Interleukin-6	↑ ↑ UA ↑	↑ ↑ ↑
Mast cell activation	\uparrow	\uparrow
T-cell activation CD+DR+ CD4+CD28–/INF+ TH1/TH2 balance	UA ↑ UA ↑ TH1 ↑	↑ ↑ TH1↑
B-cell activation	0 or ↑	0 or ↑
CRP	\uparrow	$\uparrow \uparrow$
Adhesion molecules	\uparrow	\uparrow
Endothelin	\uparrow	\uparrow
Neoangiogenesis	Ŷ	\uparrow

Source: Modified, with permission, from Pasceri and Yeh, "A tale of two diseases: atherosclerosis and rheumatoid arthritis," Circulation 1999; 100(21):2124-2126.

trast to other phenotypes of monocytes, is shown to express high levels of inflammatory cytokines such as TNF- α whereas the anti-inflammatory IL-10 is low or absent. In addition, these cells are further characterized by an upregulation of cell surface adhesion molecules, suggesting an increased capacity for cell to cell interactions.³⁰

The degree of macrophage infiltration has been shown to distinguish between unstable and stable coronary lesions. The preferential localization of macrophages in high-flow shoulder regions of the atherosclerotic plaque correlates with areas at highest risk for plaque instability. In contrast to controls, infiltrates of CD68-positive macrophages and CD3- and CD8-positive T-cells were statistically associated with the severity and frequency of superficial plaque inflammation and rupture.³¹⁻³³ This plaque instability, in part, stems from metalloproteinase (MMP-1 and MMP-2) production and release by activated macrophages within the inflamed atherosclerotic plaque.³⁴

T-Lymphocyte

Antigen-presenting macrophages induced T-cell activation and results in inflammatory amplification through T-cell release of TNF- α , and INF- γ , further activating macrophages, platelets, and smooth muscle cells.³⁵ Levels of the main specific immune markers CD4+ and CD3+/DR+ T-cells, IL-2, and IgM have all been reported to be higher in unstable than in stable angina patients.³⁶ In addition, a higher percentage of IL-2 receptor positive T-lymphocytes in culprit lesions of patients with acute coronary syndromes indicate recent activation and amplification of the immune response within plaques. These



Figure 1. Augmented platelet activity and response to agonist in patients with stable coronary artery disease versus normal controls. (Reprinted with permission from the American College of Cardiology Foundation Journal of the American College of Cardiology, 1998, vol 31, pp 352-358.)

findings support the concept that a burst of inflammatory products could initiate or accelerate the onset of an acute coronary event.³⁷

Mast cell

Mast cells have been recently identified to inhabit the vulnerable shoulder regions of the atherosclerotic plaque and to be associated with plaque erosion and rupture.^{38,39} The population size of mast cell in athrectomy tissue correlates with the clinical severity of coronary syndromes. Their presence in the adventitia of ruptured plaques has led to the postulate that histamine release may provoke coronary spasm and contributes to the onset of myocardial infarction.^{40,41} Mast cells have a primary role in the perpetuation of the inflammatory response in atherosclerosis, characterized by the production of TNF- α and neutral proteases (tryptase and chymase).^{42,43} TNF- α stimulates macrophages and smooth muscle cells to produce two prometalloproteinases-prostomelysin and procollagenase. Subsequent activation of prometalloproteinases by mast cell produced tryptase and chymase leads to fibrous cap degradation and plaque destabilization.⁴⁴

Neutrophil

As previously discussed, macrophages and T lymphocytes are the predominant cellular components of local inflammation within the atherosclerotic plaque. Neutrophils, although found sparsely in atherosclerotic plaques, play an integral part in the acute inflammatory response to tissue injury and have been implicated as a major factor in tissue damage in response to ischemia and reperfusion.⁴⁵ TNF- α , IL-8, IL-6, platelet-activating factor, and leukotrienes enhance neutrophil recruitment to ischemic and reperfused myocardium by augmenting cell adhesion molecule expression. The extent of accumulation has also been correlated to the degree of tissue injury.^{46,47} A systemic activation of neutrophils has been reported in patients with angiographically documented coronary artery disease as compared with normal controls and a subset of trauma patients providing further proof for a chronic systemic inflammatory state in patients with atherosclerosis.⁴⁸

Platelet

Traditionally, platelets have not been classified as inflammatory cells, but recent discoveries have led investigators to believe that platelets are critical constituents that tie in both inflammation and thrombosis. The presence of serologic markers of platelet activation is well established in the setting of an ACS.⁴⁹⁻⁵¹ Inflammatory cytokines induce the translocation of the cell adhesion molecule P-selectin to the surface of the platelet membrane, facilitating interactions among platelets, endothelial cells, and monocytes. Monocyte expression of tissue factor is induced by P-selectin and may be an initiator of thrombosis in areas of vascular injury.⁵²

An initial step to answer the question of whether platelet activation is a result of or results in the development of an ACS was recently reported by Furman et al⁵³ In a flow cytometric analysis patients with stable coronary artery disease were shown to not only have increased levels of circulating activated platelets with enhanced P-selectin expression, but also to have an increased propensity to form monocyte-platelet aggregates (Figure 1).⁵³ Additional evidence to implicate platelets as inflammatory mediators is the recent finding of their expression of CD40L. This transmembrane protein found on constituents of both cellular and humoral components of the inflammatory system is structurally related to TNF- α . CD40L is rapidly expressed by activated platelets and induces the expression of chemokines and cell adhesion molecules by endothelial cells thus provoking cell attraction, activation, and migration into the arterial wall.⁵⁴

MARKERS AND MEDIATORS OF INFLAMMATION C-Reactive protein (CRP)

Although many markers of inflammation have been associated with adverse cardiovascular outcome, CRP has been evaluated in every clinical phase of coronary disease. It therefore provides a superlative avenue to thoroughly discuss the prognostic significance of inflammatory mark-

ers in cardiovascular disease. CRP is an acute-phase reactant whose concentration in blood rises dramatically in response to nonspecific inflammatory stimuli. It has been convincingly linked to cardiovascular disease, initially in sera of patients after acute myocardial infarction and recently in the wall of human coronary arteries possibly linking its presence directly with the development of atherosclerosis.⁵⁵⁻⁵⁷ Whether the association reflects a casual or direct interaction, elevated levels of CRP are associated with a worse prognosis in the full spectra of atherosclerotic disease.

In the setting of a Q-wave myocardial infarction, Anzai et al⁵⁸ reported that elevated levels of CRP were associated with cardiac rupture, left ventricular aneurysm formation, and 1-year cardiac death. Even though CRP was found to be an independent predictor of these events, there remained a confounding correlation to extent of cardiac enzyme elevation in those patients without revascularization procedures.⁵⁸ Therefore, CRP levels in this study may have reflected infarct size and subsequent risk for adverse outcome.

Tommasi et al⁸ reported on the prognostic value of CRP levels in patients with a first acute myocardial infarction, uncomplicated in-hospital course, absence of residual ischemia, and normal left ventricular function. Only increased CRP levels were independently associated to the incidence of patients who developed cardiac events (cardiac death, new-onset angina, and recurrent myocardial infarction) (Figure 2).⁸ Importantly, there was no correlation between CRP levels and extent of rise of cardiac enzymes.

Although numerous studies have shown that an elevated CRP in the setting of unstable angina and non-Qwave myocardial infarction is associated with worse prognosis,^{6,7,59-61} Biassicci et al⁶² reported on the prognostic significance of CRP elevation in patients with unstable angina without myocardial injury. They excluded those patients with elevated levels of cardiac enzymes at entry to avoid the interplay of myocardial necrosis on CRP and future events. They reported that an elevated discharge CRP was strongly associated with recurrent coronary instability and myocardial infarction (Figure 3) and, interestingly, 42% of patients had persistent elevation of CRP 3 months after hospital discharge. Adjunctive evidence that elevated CRP levels possess predictive power exceeding their association with myonecrosis is their independent and additive prognostic value to markers of myocardial injury, such as troponin T and I.63,64

In the Thrombolysis in Myocardial Infarction (TIMI) IIA trial, a dose-ranging trial for enoxaparin in UA and NQMI, elevated CRP correlated with increased 14-day mortality (Figure 4). Most importantly, these findings existed even in patients with a negative rapid troponin T assay, thereby dissociating myonecrosis from CRP's prognostic power.⁶⁴ Milazzo et al⁶⁵ reported that in patients undergoing CABG a preoperative elevation of CRP has prognostic significance (Figure 5). CRP levels <3 mg/L and \geq 3 mg/L were associated with new ischemic events in 4% vs. 25% of patients, respectively.

In the setting of percutaneous coronary revasculariza-

tion, a hyperresponsive reaction of the inflammatory system, defined by elevation of CRP, IL-6, and serum amyloid A after angioplasty, was recently presumed to portend a worse prognosis.⁶⁶ Gaspardone et al⁶⁷ confirmed this by showing a persistent elevation in CRP 72 hours after coronary artery stenting (excluding patients with periprocedural myocardial infarction) pinpointed all patients who later suffered an adverse outcome. In contrast, no cardiac events occurred in those with normal levels at 1 year follow-up (**Figure 6**).

Ex vivo studies have recently introduced the concept that detecting heat release by inflammatory cells within an atherosclerotic plaque may predict future instability and rupture.⁶⁸ Stefanadis et al,⁶⁹ using a thermography catheter, demonstrated heterogeneity in heat production of 20%, 40%, and 67% in atherosclerotic plaques of patients with stable angina, unstable angina, and acute my-ocardial infarction, respectively. Most importantly there was a significant correlation between thermal heterogeneity and baseline CRP (Figure 7).⁶⁹

More conclusive evidence that chronic, indolent inflammation plays a principal role in the development and progression of atherosclerosis has come from the longterm follow-up of patients with no known atherosclerotic disease but increased levels of CRP. Among 14,916 apparently healthy men participating in the Physician's Health Study an elevated level of high-sensitivity CRP (HsCRP), which detects CRP levels as low as 0.175 mg/L, added to the predictive value of elevated lipids in predicting first myocardial infarction (Figure 8).⁷⁰ Similarly, in the Women's Health Study, those who developed cardiovascular events had higher baseline CRP levels than control subjects, with the highest levels at baseline being associated with a five- and seven-fold increase in any vascular event and combined stroke or myocardial infarction, respectively.⁷¹

Additional evidence that CRP levels are strong predictors of future cardiac events in apparently healthy men was recently published from the Monitoring Trends and Determinants in Cardiovascular Disease Study (MONICA). Patients in the highest quintile of CRP level had a 2.6fold increased risk of suffering a fatal or nonfatal myocardial infarction or sudden cardiac death.⁷² These findings strongly support the pivotal role that inflammation plays in the destabilization of atherosclerosis.

The question remains what if any direct role CRP plays in the development of atherosclerosis. A possible explanation supporting CRP as an indirect cardiovascular risk factor is that it reflects inflammation related to coronary vessel pathogens, extent of atherosclerosis, myocardial necrosis, myocardial ischemia, and activity of circulating proinflammatory cytokines.⁷³ Direct evidence for CRP's role in the pathogenesis of atherosclerosis is that its presence in the arterial wall predicts severity of atherosclerosis and that it is able to bind to damaged membranes and lipids, activate complement, and stimulate production of tissue factor from activated macrophages.⁷⁴⁻⁷⁷ Irrespective of its pathologic role, there is overwhelming evidence that CRP, a sensitive marker of inflammation, is a powerful predictor of future cardiac events in patients with Q-wave



Figure 2. (Left) The event-free survival with respect to level of CRP in patients after an uncomplicated myocardial infarction. (Right) The distribution of events per quartile of CRP elevation. (Reprinted from the American Journal of Cardiology, vol 83, Tommasi et al, "C-reactive protein as marker for cardiac ischemic events in the year after a first, uncomplicated myocardial infarction," pp 1595-1599, Copyright 1999, with permission from Excerpta Medica.)



Figure 3. Cumulative event-free survival in patients with unstable angina, negative cardiac enzymes, and elevated discharge CRP. (Reprinted, with permission, from Biasucci et al, "Elevated levels of C-reactive protein at discharge in patients with unstable angina predict recurrent instability," Circulation 1999; 99(7):855-860.)

MI, non-Q MI, unstable angina, stable angina, in patients who have undergone CABG and percutaneous coronary stenting, and recently in apparently healthy men and women. Recently the FDA has approved the use of a high-sensitivity CRP assay (Behring) as a prognostic test in the evaluation of patients with or expected atherosclerosis. This test has been most studied in those patients without clinically apparent atherosclerosis as a predictor of future cardiovascular events based on tertile of elevation.

Tumor necrosis factor- α (TNF- α) and other mediators

TNF- α is a pleiotropic proinflammatory cytokine with a wide range of effects that extend across a spectrum of pathologic conditions. Present in atherosclerotic lesions,⁷⁸ TNF- α appears to be one of the most important influences on the progression of atherosclerosis. Its upregulation is known to mediate and amplify a multitude of



Figure 4. Independent and additive predictive value of CRP and cTNT (early positive defined by being positive in <10 minutes) on 14-day mortality. (Reprinted with permission from the American College of Cardiology Foundation Journal of the American College of Cardiology, 1998, vol 31, pp 1460-1465.)

interactions resulting in progressive inflammation, plaque destabilization, and prothrombotic tendencies⁷⁹⁻⁸⁷ (**Table 2**). Treatment with a chimeric mAb to TNF- α has been shown to suppress inflammation and improve patient well-being in rheumatoid arthritis. Administration of anti-TNF- α Ab was recently shown to rapidly downregulate a spectrum of cytokines (IL-6), cytokine inhibitors (TNF receptors p75 and p55), and acute-phase proteins (amyloid A, haptoglobin, and fibrinogen).⁸⁸ This potent suppression of markers and mediators of inflammation may have tremendous potential in preventing progression of atherosclerosis.

IL-6 and IL-1 Ra (IL-1 receptor antagonist) not only have been shown to be elevated in the setting of ACS, but also are associated with increased risk of in-hospital events.⁸⁹ IL-6, produced by a variety of inflammatory cell types, has been shown to remain elevated up to 4 weeks after a myocardial infarction. Its properties increase fib-



Figure 5. Cumulative proportion of ischemic events in patients with elevated CRP prior to coronary artery bypass grafting. (Reprinted from the American Journal of Cardiology, vol 84, Milazzo et al, "Elevated levels of C-reactive protein before coronary artery bypass grafting predict recurrence of ischemic events," pp 459-461, Copyright 1999, with permission from Excerpta Medica.)



Figure 7. The correlation between thermal heterogeneity and level of CRP in control, stable angina, unstable angina, and acute myocardial infarction patients. (Reprinted, with permission, from Stefanadis et al, "Thermal heterogeneity within human atherosclerotic coronary arteries detected in vivo," Circulation 1999; 99(15):1965-1971.)

rinogen and PAI-1, promote adhesion of neutrophils and myocytes during myocardial reperfusion, and produce a negative inotropic effect on the myocardium.90-94 Pannitteri et al⁹⁵ reported that levels of IL-8 not only are elevated in the setting of acute myocardial infarction but that they precede the levels of IL-6 and parallel the kinetics of CPK. IL-8 is a powerful trigger for firm adhesion of monocytes to vascular endothelium, may play a potential atherogenic role by inhibiting local inhibitors of metalloproteinases in atherosclerotic plaques, and stimulates smooth muscle cell migration.^{96,97} IL-4 and IL-13 have been shown to enhance the ability of activated human monocytes to oxidize LDL, thus potentiating its toxic effects.98 OxLDL induces IFN-y production by T-helper-1-like cells, which are known to inhibit local collagen synthesis by SMC, stimulate expression of tissue factor and CD40, and selectively induce MCP-1.98-100 Many other cytokines have been implicated in immunity, inflammation, thrombosis, and angiogenesis.¹⁰¹

The above discussion underscores the vast trafficking,



Figure 6. Event-free survival with respect to persistent elevation of CRP >72 hours after coronary stenting. (Reprinted from the American Journal of Cardiology, vol 82, Gaspardone et al, "Predictive value of C-reactive protein after successful coronary-artery stenting in patients with stable angina," pp 515-518, Copyright 1998, with permission from Excerpta Medica.)



Figure 8. Predictive value for lipoprotein(a), total homocysteine, total cholesterol, fibrinogen, t-PA antigen, ratio of total cholesterol to HDL, CRP, and CRP plus total cholesterol HDL ratio. (From Ridker PM et al, Ann Intern Med 1999; 130:933-937, with permission.)

redundancy, and interplay of the cytokine system. Each mediator, though, must work through specific receptors and ultimately regulate gene expression of proteins vital to the potentiation and regulation of the inflammatory cascade. Nuclear factor κ -B (NF- κ B), peroxisome proliferator-activated receptor activators (PPARs), CD40 receptor and its-ligand, and inducible cyclooxygenase enzyme (Cox-2) are avidly being investigated as we attempt to discover the final common pathway of inflammation and its role in atherosclerosis.

NF-κB

NF- κ B is a transcription factor located in the cytoplasm of many cells as an inactive complex associated with a specific class of inhibitory proteins, called I κ B. This complex binds and prevents nuclear translocation and DNA binding of NF- κ B.¹⁰² In response to inflammatory stimuli I κ B is eventually degraded and NF- κ B is released and transported to the nucleus. In the nuclei, NF- κ B can initiate or regulate early response gene transcription by binding to promotor or enhancer regions.¹⁰³ NF-

TABLE 2

Proinflammatory and thrombotic properties of $\mathsf{TNF-}\alpha$

nflammatory properties
Regulation of macrophage colony-stimulating factor
Regulation of cell adhesion molecules
Modulation of smooth muscle cell phenotype
Induction of IL-1 mRNA
Inhibition of endothelial cell apoptosis
Plague destabilization
Induces smooth muscle cell interstitial collagenases
Neutral effect on tissue inhibitors of
metalloproteinases

- Thrombotic properties
 - Augments transcription and expression of tissue factor Decreases in activity of thrombomodulin-C and
 - tissue-type plasminogen activator Increases production of plasminogen activator inhibitor
 - Increases release of Von Willebrand factor

 κ B is known to regulate or be regulated by genes involved in every aspect of the proinflammatory cascade.^{104,105} TNF-α and IL-1 are two important inducers, contributing to a positive feedback loop for NF-κB activation. As a consequence, there is a continuous upregulation of cytokines and perpetuation of inflammation.¹⁰³ NF-κB has been implicated in a variety of inflammatory diseases, such as allograft rejection, rheumatoid arthritis (RA), asthma, and inflammatory bowel disease.¹⁰⁴ In RA, NFκB is overly expressed in synovial tissue, associated with surface expression of cell adhesion molecules, production of cytokines, and upregulation of the inducible isoform of cyclo-oxygenase (Cox-2). These processes are parallel to those found in atherosclerotic lesions.¹⁰⁴

NF-KB activity is enhanced by known cardiac risk factors such as very low-density lipoprotein, OxLDL, hyperglycemia, and elevated levels of angiotensin II. On the contrary, its activity is inhibited by HMG-CoA reductase inhibitors, antioxidants, and gallates (phenolic compounds found abundantly in red wine).¹⁰⁶⁻¹¹¹ Recently, Ritchie¹¹² reported data showing that NF- κ B is activated in patients with unstable angina without evidence of myonecrosis and is therefore potentially linked in plaque disruption. Immunosuppression with glucocorticoids, gold, cyclosporin, FK506, and, importantly, aspirin and salicylates is known to inhibit NF- κ B. Kopp et al¹¹³ demonstrated that aspirin inhibits NF- κ B activity by preventing the degradation of I κ B, while Weber et al¹¹⁴ established aspirin's ability to inhibit TNF- α -stimulated NF- κ B activity.

CD40 and CD40L

CD40 is a phosphorylated 49-kDa glycoprotein expressed on B-lymphocytes, fibroblasts, monocytes, platelets, epithelial cells, and endothelial cells.¹¹⁵ CD40L, also named CD154 or gp39, belongs to the TNF family of cytokines. The presence of CD40 and CD40L has been



Figure 9. Levels of soluble CD40L in normal controls, stable angina, and unstable angina patients. (Reprinted, with permission, from Aukrust et al, "Enhanced levels of soluble and membrane-bound CD40 ligand in patients with unstable angina," Circulation 1999; 100(6):614-620.)

found in human atheroma, and their association is implicated with expression of cell adhesion molecules, cytokines, matrix metalloproteinases, and tissue factor.54,115 Anti-CD40L has been shown to regulate autoimmune diseases such as lupus nephritis, skin and cardiac allograft rejection, and multiple sclerosis in experimental models.¹¹⁶⁻¹¹⁸ Mach et al¹¹⁹ reported a reduction in aortic atherosclerotic lesion size, fewer T-lymphocytes and macrophages, and a decreased presence of cell adhesion molecules in atheroma in cholesterol-fed mice lacking the LDL receptor when treated with anti-CD40L antibody.¹¹⁹ Aukrust et al¹²⁰ recently reported elevated levels of CD40-CD40L in patients with angina pectoris. Patients with unstable angina had significantly higher levels than those with stable angina, allowing the authors to conclude that presence of CD40-CD40L may have a pathologic role in plaque destabilization and the development of ACS¹²⁰ (Figure 9).

PPAR

Peroxisomal proliferator-activated receptors (PPARs), including PPAR- α , PPAR- γ , and PPAR- δ , are a group of nuclear transcription factors playing a key role in adipogenesis and lipid metabolism.¹²¹ Recently, modulation of the development and progression of atherosclerosis has been substantiated by research that appears to link PPAR activity with the regulation of inflammation and plaque stability by their interactions with macrophages, endothelial cells, smooth muscle cells, and metalloproteinases. Ricote et al¹²² found PPAR- γ to be upregulated in activated macrophages and to inhibit gelatinase B, nitric oxide synthase, and scavenger receptors. OxLDL has been shown to induce PPAR- γ expression in macrophages, resulting in monocyte differentiation and enhanced uptake in OxLDL.^{123,124} Max et al¹²⁵ recently reported elevated levels of PPAR- γ expression on monocytes in human atherosclerotic lesions as compared to normal controls. Furthermore, PPAR- γ stimulation leads to a concentration-dependent decrement in monocyte-derived metalloproteinase activity. Finally, PPAR- α and - γ have been implicated in the induction of macrophage apoptosis through

SII-136 CLEVELAND CLINIC JOURNAL OF MEDICINE

VOLUME 69 • SUPPLEMENT II

Downloaded from www.ccjm.org on June 3, 2025. For personal use only. All other uses require permission.



Figure 10. Relative benefit of ASA with respect to quartile of CRP. Data are shown allocated to ASA (open bars) and placebo (solid bars). (From Ridker et al, "Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men," N Engl J Med 1997; 336:973-979, with permission. Copyright © 1997 Massachusetts Medical Society. All rights reserved.)

inhibition of NF- κ B antiapoptotic pathways.¹²⁶ Endothelial cells also appear to be under the influence of PPARs by the regulation of leukocyte/endothelial cell interactions. Jackson et al¹²⁷ demonstrated an inhibitory effect of stimulated PPAR on endothelial cell expression of VCAM-1. In addition, stimulated PPAR- α has been shown to inhibit TNF- α -mediated endothelial cell VCAM-1 expression, COX-2 expression, IL-1 induced production of IL-6, and thrombin-induced endothelial-1 production.¹²⁸⁻¹³⁰

Key stimulatory PPAR ligands are naturally occurring prostaglandins, as well as synthetic antidiabetic and antilipidemic drugs. Gemfibrozil, a finofibrate and stimulator of PPAR- α , has recently been shown to dramatically reduce IL-1-induced production of IL-6, expression of COX-2 in human smooth muscle cells, and cardiovascular events in patients with low HDL levels. Importantly, this reduction in cardiovascular events was independent of LDL levels.^{130,131} Troglitazone, an insulin sensitizer and PPAR- γ ligand, demonstrates a range of anti-inflammatory and potential plaque-stabilizing activities such as PPAR- γ -induced inhibition of macrophage metalloproteinases.¹²⁵

Currently, the complex activities of PPARs and their ligands are not completely understood, although ligands with positive effects on lipid lowering (finofibrates) and glycemic control (troglitazone) would suggest that these transcriptional factors are clinically beneficial and mainly antiatherogenic.

Cyclo-oxygenase-2 (COX-2)

There are two distinct isoforms of cyclo-oxygenase (COX-1 and COX-2). These enzymes are necessary in the conversion of arachidonic acid to prostaglandin G2 and H2, which are potent agonists to the inflammatory cascade.^{132,133} The ability of ASA and other NSAIDs to inhibit inflammation through their regulation of COX-1 was first described by Vane in 1971.¹³² It was not until



Figure 11. Cardiovascular events allocated to background of NSAID (solid bars) and control (open bars).

1991 that an inducible form of the COX enzyme was discovered, COX-2.134 Although a weak COX-2 inhibitor, aspirin and most available NSAIDs by virtue of their preferential COX-1 inhibitory effects, provide minimal antiinflammatory action at doses not associated with significant side effects. COX-2 receptors are scantly expressed in the gastrointestinal tract or platelets and therefore likely provide augmented inflammatory control with few adverse effects.¹³⁵ COX-2 is felt to be the principal isoform that participates in inflammation and has been recently found to be widely expressed in atherosclerotic tissue.136,137 Macrophage COX-2 mRNA expression has been shown to be induced by inflammatory cytokines such as INF- γ , TNF- α , and lipopolysaccharide, while other cytokines with anti-inflammatory properties, such as IL-10, have been shown to inhibit its induction.^{138,139}

Speir et al¹⁴⁰ recently demonstrated a reduction in reactive oxygen species generation in CMV infected smooth muscle cells when pretreated with NSAIDs. This reduction was thought mainly to be due to inhibition of the COX-2 enzyme.¹⁴⁰ Although most investigations have described COX-2 as a proinflammatory mediator, recent reports by Cockerill et al141 and Bishop-Bailey et al¹⁴² have provided evidence for its anti-inflammatory potential. They demonstrated that HDL enhanced the expression of COX-2-dependent prostaglandin-I2, which is known to inhibit platelet and leukocyte activity. In addition, inhibition of IL-1- β resulted in upregulation of COX-2 and downregulation of the cell adhesion molecule ICAM-1. Although questions still remain, anti-inflammatory treatment such as aspirin, with its unquestionable beneficial effects, and a recent retrospective analysis of NSAIDs in patients after myocardial infarction that demonstrated a reduction in cardiac mortality and adverse events (Figure 10),¹⁴³ suggest the potential for augmented clinical benefit with more potent and selective cyclooxygenase inhibition.

THE FUTURE OF INFLAMMATION CONTROL IN ACS

Aspirin, initially thought of mainly as an antiplatelet drug in the battle with atherosclerotic heart disease, is becoming more recognized for its anti-inflammatory properties. In addition to aspirin's COX-1 and weak COX-2 activity,



Figure 12. (Left) CRP at baseline and 5 years for patients treated with pravastatin or placebo. (Right) The change in CRP according to LDL level. Data are shown allocated to pravastatin (open bars) and placebo (solid bars). (Reprinted, with permission, from Ridker et al, "Long-term effects of pravastatin on plasma concentration of C-reactive protein," Circulation 1999; 100(3):230-235.)



Figure 13. Cardiovascular endpoints allocated by patients receiving ramipril (open bars) or placebo (solid bars).

the inhibition of NF- κ B activity is achieved by inhibiting both the degradation of I κ B and effects of TNF- α . Clinical evidence to support aspirin's anti-inflammatory role has been reported by Ridker et al.¹⁴⁴ Aspirin reduced first MI in the Physicians Health Study, and this effect was directly related to the baseline CRP level¹⁴⁴ (Figure 11). In addition, the recent negative results of the oral IIb/IIIa receptor inhibitor may be explained in part by the lack of aspirin's anti-inflammatory properties in the group receiving sole oral IIb/IIIa treatment.

A paradigm shift in thought may be evolving in favor of the anti-inflammatory properties of aspirin being more salient than its relatively weak antiplatelet effects in the reduction of ischemic cardiac events. HMG-CoA reductase inhibitors have been shown to dramatically reduce cardiovascular mortality and morbidity although the reduction in events is not linear with the reduction of LDL cholesterol below 125 mg/dL.¹⁴⁵ In an analysis of the Cholesterol and Recurrent Events (CARE) trial, Ridker et al¹⁴⁶ reported a significant 22% drop in CRP over a 5year period in those treated with pravastatin versus placebo. Interestingly, CRP rose even in the placebo-treated arm which realized a reduction in LDL cholesterol

(Figure 12). Evidence continues to mount suggesting an anti-inflammatory role for HMG-CoA reductase inhibitors as these agents have been shown to alter regulation of DNA transcription, regulate natural-killer-cell cytotoxicity, inhibit platelet-derived growth factor-induced DNA synthesis, and decrease macrophage production of metalloproteinases.¹⁴⁶⁻¹⁴⁹ ACE inhibitors have recently been demonstrated to possess potent anti-inflammatory properties that may explain their regulating effects on atherosclerotic driven endpoints. ACE inhibitors have been shown to exhibit antiproliferative and antimigratory effects on SMC and leukocytes, restore endothelial function, modulate platelet effects, and promote endogenous $fibrinolysis. ^{150} \\$ The Heart Outcomes Prevention Evaluation (HOPE) study, a study of patients with vascular disease and no known heart failure, reported a dramatic and significant decrease in cardiovascular death, MI, and stroke in patients treated with ramipril versus placebo (Figure 13).¹⁵¹ Finofibrates and insulin sensitizers such as troglitazone are stimulators of PPAR receptors and are currently receiving attention for their anti-inflammatory and antiatherogenic potential. Unfortunately, not all methods of inflammatory control have realized a positive clinical outcome. Prevention of reperfusion injury in patients presenting with ACS by inhibiting leukocyte adhesion was recently reported from the HALT MI study. There was no significant reduction in infarct size and unfortunately a significant increase in infection rates in those randomized to high dose of the CD11/CD18 inhibitor.¹⁵² This trial underscores the careful balance needed between adequate anti-inflammatory control and clinically significant immunosuppression.

Even though CRP and HsCRP have been shown to predict risk of future adverse cardiovascular events in virtually all patient subgroups, treatment options are limited to drugs not specifically heralded for their anti-inflammatory properties. Novel downstream approaches with the use of TNF- α , CD40L, NF- κ B, and COX-2 inhibitors are under in vitro and animal investigations to

determine their potential role in the battle against atherosclerosis. Treatment of atherosclerosis as an inflammatory disease should first focus on those pathogens known to initiate and propagate this disease, such as hypercholesterolemia, hypertension, diabetes, hyperhomocysteinemia, smoking, and possible infection. The second ap-

REFERENCES

- 1. Alexander RW. Inflammation and coronary artery disease. N Engl J Med 1994; 331:468–469.
- Ross R, Glomset JA. The pathogenesis of atherosclerosis (first of two parts). N Engl J Med 1976; 295:369–377.
- Ross R. The pathogenesis of atherosclerosis—an update. N Engl J Med 1986; 314:488–500.
- Ross R. The pathogenesis of atherosclerosis: a perspective for the 1990s. Nature 1993; 362:801–809.
- Ross R. Atherosclerosis—an inflammatory disease. N Engl J Med 1999; 340:115–126.
- Toss H, Lindahl B, Siegbahn A, Wallentin L. Prognostic influence of increased fibrinogen and C-reactive protein levels in unstable coronary artery disease. FRISC Study Group. Fragmin during instability in coronary artery disease. Circulation 1997; 96:4204–4210.
- Liuzzo G, Biasucci LM, Gallimore JR, Grillo RL, Rebuzzi AG, Pepys MB, Maseri A. The prognostic value of C-reactive protein and serum amyloid a protein in severe unstable angina. N Engl J Med 1994; 331:417–424.
- Tommasi S, Carluccio E, Bentivoglio M, Buccolieri M, Marioti M, Politano M, Corea L. C-reactive protein as marker for cardiac ischemic events in the year after a first, uncomplicated myocardial infarction. Am J Cardiol 1999; 83:1595–1599.
- Biasucci LM, Liuzzo G, Caligiuri G, Quaranta G, Andreotti F, Sperti G, van de Greef W, Rebuzzi AG, Kluft C, Maseri A. Temporal relation between ischemic episodes and activation of the coagulation system in unstable angina. Circulation 1996; 93:2121–2127.
- Bhagat K. Endothelial function and myocardial infarction. Cardiovasc Res 1998; 39:312–317.
- Kinlay S, Selwyn AP, Libby P, Ganz P. Inflammation, the endothelium, and the acute coronary syndromes. J Cardiovasc Pharmacol 1998; 32:S62–S66.
- 12. Noll G, Luscher TF. The endothelium in acute coronary syndromes. Eur Heart J 1998; 19(suppl C):C30–C38.
- Gonzalez-Amaro R, Diaz-Gonzalez F, Sanchez-Madrid F. Adhesion molecules in inflammatory diseases. Drugs 1998; 56:977–988.
- 14. Petruzzelli L, Takami M, Humes HD. Structure and function of cell adhesion molecules. Am J Med 1999; 106:467–476.
- Frenette PS, Wagner DD. Adhesion molecules—part I. N Engl J Med 1996; 334:1526–1529.
- Frenette PS, Wagner DD. Adhesion molecules—part II: blood vessels and blood cells. N Engl J Med 1996; 335:43–45.
- 17. Springer TA. Adhesion receptors of the immune system. Nature 1990; 346:425–434.
- Braun M, Pietsch P, Schror K, Baumann G, Felix SB. Cellular adhesion molecules on vascular smooth muscle cells. Cardiovasc Res 1999; 41:395–401.
- Marx N NF-J, Fischer A, Heimerl S, Dickfeld T. Induction of monocyte procoagulant activity by adhesion on vascular smooth muscle cells and ICAM-1 transfected CHO-cells. Circulation 1997; 96(suppl):I–112.
- Phillips DR, Charo IF, Parise LV, Fitzgerald LA. The platelet membrane glycoprotein IIb-IIIa complex. Blood 1988; 71:831–843.
- De caterina R BG, Lazzerini G, Dell'Omo G, Petrucci R, Morale M, Carmassi F, Pedrinelli R. Soluble vascular cell adhesion molecule-1 as a biohumoral correlate of athersclerosis. Arterioscler Thromb Vasc Biol 1997; 17:2646–2654.
- Ridker PM, Hennekens CH, Roitman-Johnson B, Stampfer MJ, Allen J. Plasma concentration of soluble intercellular adhesion molecule 1 and risks of future myocardial infarction in apparently healthy men. Lancet 1998; 351:88–92.

proach should be to uncover the etiology of the nearly 50% of patients who present with an ACS without known cardiac risk factors. Finally, further investigation is needed to determine the clinical efficacy of adjunctive anti-inflammatory therapy on the background of pathogen directed treatment.

- Rohde LE, Hennekens CH, Ridker PM. Cross-sectional study of soluble intercellular adhesion molecule-1 and cardiovascular risk factors in apparently healthy men. Arterioscler Thromb Vasc Biol 1999; 19:1595–1599.
- 24. Tenaglia AN, Buda AJ, Wilkins RG, Barron MK, Jeffords PR, Vo K, Jordan MO, Kusnick BA, Lefer DJ. Levels of expression of P-selectin, E-selectin, and intercellular adhesion molecule-1 in coronary atherectomy specimens from patients with stable and unstable angina pectoris. Am J Cardiol 1997; 79:742–747.
- Davis LS, Kavanaugh AF, Nichols LA, Lipsky PE. Induction of persistent T cell hyporesponsiveness in vivo by monoclonal antibody to ICAM-1 in patients with rheumatoid arthritis. J Immunol 1995; 154:3525–3537.
- Buerke M, Weyrich AS, Zheng Z, Gaeta FC, Forrest MJ, Lefer AM. Sialyl Lewisx-containing oligosaccharide attenuates myocardial reperfusion injury in cats. J Clin Invest 1994; 93:1140–1148.
- Silver MJ, Sutton JM, Hook S, Lee P, Malycky JL, Phillips ML, Ellis SG, Topol EJ, Nicolini FA. Adjunctive selectin blockade successfully reduces infarct size beyond thrombolysis in the electrolytic canine coronary artery model. Circulation 1995; 92:492–499.
- Gown AM, Tsukada T, Ross R. Human atherosclerosis. II. Immunocytochemical analysis of the cellular composition of human atherosclerotic lesions. Am J Pathol 1986; 125:191–207.
- Schmitz G, Herr AS, Rothe G. T-lymphocytes and monocytes in athergenesis. Herz 1998; 23:168–177.
- Frankenberger M, Sternsdorf T, Pechumer H, Pforte A, Ziegler-Heitbrock HW. Differential cytokine expression in human blood monocyte subpopulations: a polymerase chain reaction analysis Blood 1996; 87:373–377.
- Boyle JJ. Association of coronary plaque rupture and atherosclerotic inflammation. J Pathol 1997; 181:93–99.
- Moreno PR, Falk E, Palacios IF, Newell JB, Fuster V, Fallon JT. Macrophage infiltration in acute coronary syndromes. Implications for plaque rupture. Circulation 1994; 90:775–778.
- Dirksen MT, van der Wal AC. van den Berg FM, van der Loos CM, Becker AE. Distribution of inflammatory cells in atherosclerotic plaques relates to the direction of flow. Circulation 1998; 98:2000–2003.
- Shah PK, Falk E, Badimon JJ, Fernandez-Ortiz A, Mailhac A, Villareal-Levy G, Fallon JT, Regnstrom J, Fuster V. Human monocyte-derived macrophages induce collagen breakdown in fibrous caps of atherosclerotic plaques. Potential role of matrix-degrading metalloproteinases and implications for plaque rupture. Circulation 1995; 92:1565–1569.
- Hansson GK, Jonasson L, Seifert PS, Stemme S. Immune mechanisms in atherosclerosis. Arteriosclerosis 1989; 9:567–578.
- Caligiuri G, Liuzzo G, Biasucci LM, Maseri A. Immune system activation follows inflammation in unstable angina: pathogenetic implications. J Am Coll Cardiol 1998; 32:1295–1304.
- Van der Wal AC, Piek JJ, de Boer OJ, Koch KT, Teeling P, Van der Loos CM, Becker AE. Recent activation of the plaque immune response in coronary lesions underlying acute coronary syndromes. Heart 1998; 80:14–18.
- Kaartinen M, Penttila A, Kovanen PT. Accumulation of activated mast cells in the shoulder region of humanl coronary atheroma, the predilection site of atheromatous rupture Circulation 1994; 90:1669–1678.
- Kovanen PT, Kaartinen M, Paavonen T. Infiltrates of activated mast cells at the site of coronary atheromatous erosion or rupture in myocardial infarction. Circulation 1995; 92:1084–1088.
- Kaartinen M, van der Wal AC, van der Loos CM, Piek JJ, Koch KT, Becker AE, Kovanen PT. Mast cell infiltration in acute coronary syndromes: implications for plaque rupture. J Am Coll

Cardiol 1998; 32:606-612.

- Laine P, Kaartinen M, Penttila A, Panula P, Paavonen T, Kovanen PT. Association between myocardial infarction and the mast cells in the adventitia of the infarct-related coronary artery. Circulation 1999; 99:361–369.
- Irani AA, Schechter NM, Craic SS, DeBlois G, Schwartz LB. Two types of human mast cells that have distinct neutral protease compositions. Proc Natl Acad Sci USA 1986; 83:4464–4468.
- Kaartinen M, Penttila A, Kovanen PT. Mast cells in rupture-prone areas of human coronary atheromas produce and store TNF-alpha. Circulation 1996; 94:2787–2792.
- Hibbs MS, Hoidal JR, Kang AH. Expression of a metalloproteinase that degrades native type V collagen and denatured collagens by cultured human alveolar macrophages. J Clin Invest 1987; 80:1644–1650.
- 45. Takeshita S, Isshiki T, Ochiai M, Ishikawa T, Nishiyama Y, Fusano T, Toyoizumi H, Kondo K, Ono Y, Sato T. Systemic inflammatory responses in acute coronary syndrome: increased activity observed in polymorphonuclear leukocytes but not T Iymphocytes. Atherosclerosis 1997; 135:187–192.
- Dreyer WJ, Smith CW, Michael LH, Rossen RD, Hughes BJ, Entman ML. Anderson DC. Canine neutrophil activation by cardiac lymph obtained during reperfusion of ischemic myocardium. Circ Res 1989; 65:1751–1762.
- Smith EFd, Egan JW, Bugelski PJ, Hillegass LM, Hill DE, Griswold DE. Temporal relation between neutrophil accumulation and myocardial reperfusion injury. Am J Physiol 1988; 255:H1060–H1068.
- 48. Kassirer M, Zeltser D, Prochorov V, Schoenman G, Frimerman A, Keren G, Shapira I, Miller H, Roth A, Arber N, Eldor A, Berliner S. Increased expression of the CDIIb/CD18 antigen on the surface of peripheral white blood cells in patients with ischemic heart disease: further evidence for smoldering inflammation in patients with atherosclerosis. Am Heart J 1999; 138:555–559.
- Fuster V, Badimon L, Badimon JJ, Chesebro JH. The pathogenesis of coronary artery disease and the acute coronary syndromes (1). N Engl J Med 1992; 326:242–250.
- Fuster V, Badimon L, Badimon JJ, Chesebro JH. The pathogenesis of coronary artery disease and the acute coronary syndromes (2). N Engl J Med 1992; 326:310–318.
- Fitzgerald DJ, Roy L, Catella F, FitzGerald GA. Platelet activation in unstable coronary disease. N Engl J Med 1986; 315:983–989.
- Celi A, Pellegrini G, Lorenzet R, De Blasi A, Ready N, Furie BC, Furie B. P-selectin induces the expression of tissue factor on monocytes. Proc Natl Acad Sci USA 1994; 91:8767–8771.
- 53. Furman MI, Benoit SE, Barnard MR, Valeri CR, Borbone ML, Becker RC, Hechtman HB, Michelson AD. Increased platelet reactivity and circulating monocyte-platelet aggregates in patients with stable coronary artery disease. J Am Coll Cardiol 1998; 31:352–358.
- Henn V, Slupsky JR, Grafe M, Anagnostopoulos I, Forster R, Muller-Berghaus G, Kroczek RA. CD40 ligand on activated platelets triggers an inflammatory reaction of endothelial cells. Nature 1998; 391:591–594.
- Zimmermann-Gorska I, Kujawa H, Drygas J. Studies of acute phase reactants in myocardial infarction. Pol Med J 1972; 11:779–785.
- Jain VC. An evaluation of C-reactive protein test in acute myocardial infarction. Indian Heart J 1968; 20:16–21.
- Zhang YX, Cliff WJ, Schoefl Gl, Higgins C. Coronary C-reactive protein distribution: its relation to development of atherosclerosis. Atherosclerosis 1999; 145:375–379.
- Anzai T, Yoshikawa T, Shiraki H, Asakura Y, Akaishi M, Mitamura H, Ogawa S. C-reactive protein as a predictor of infarct expansion and cardiac rupture after a first Q-wave acute myocardial infarction. Circulation 1997; 96:778–784.
- Haverkate F, Thompson SG, Pyke SD, Gallimore JR, Pepys MB. Production of C-reactive protein and risk of coronary events in stable and unstablc angina. European Concerted Action on Thrombosis and Disabilities Angina Pectoris Study Group. Lancet 1997; 349:462–466.
- 60. Verheggen PW, de Maat MP, Cats VM, Haverkate F, Zwinderman AH, Kluft C, Bruschke AV. Inflammatory status as a main determinant of outcome in patients with unstable angina, independent of coagulation activation and endothelial cell function. Eur Heart

J 1999; 20:567–574.

- Biasucci LM, Vitelli A, Liuzzo G, Altamura S, Caligiuri G, Monaco C, Rebuzzi AG, Ciliberto G, Maseri A. Elevated levels of interleukin-6 in unstable angina. Circulation 1996; 94:874–877.
- Biasucci LM, Liuzzo G, Grillo RL, Caligiuri G, Rebuzzi AG, Buffon A, Summaria F, Ginnetti F, Fadda G, Maseri A. Elevated levels of C-reactive protein at discharge in patients with unstable angina predict recurrent instability. Circulation 1999; 99:855–860.
- 63. Rebuzzi AG, Quaranta G, Liuzzo G, Caligiuri G, Lanza GA, Gallimore JR, Grillo RL, Cianflone D, Biasucci LM, Maseri A. Incremental prognostic value of serum levels of troponin T and Creactive protein on admission in patients with unstable angina pectoris. Am J Cardiol 1998; 82:715–719.
- 64. Morrow DA, Rifai N, Antman EM, Weiner DL, McCabe CH, Cannon CP, Braunwald E. C-reactive protein is a potent predictor of mortality independently of and in combination with troponin T in acute coronary syndromes: a TIMI 11A substudy. Thrombolysis in Myocardial Infarction. J Am Coll Cardiol 1998; 31:1460–1465.
- Milazzo D, Biasucci LM, Luciani N, Martinelli L, Canosa C, Schiavello R, Maseri A, Possati G. Elevated levels of C-reactive protein before coronary artery bypass grafting predict recurrence of ischemic events. Am J Cardiol 1999; 84:459–461, A9.
- Liuzzo G, Buffon A, Biasucci LM, Gallimore JR, Caligiuri G, Vitelli A, Altamura S, Ciliberto G, Rebuzzi AG, Crea F, Pepys MB, Maseri A. Enhanced inflammatory response to coronary angioplasty in patients with severe unstable angina. Circulation 1998; 98:2370–2376.
- Gaspardone A, Crea F, Versaci F, Tomai F, Pellegrino A, Chiariello L, Gioffre PA. Predictive value of C-reactive protein after successful coronary-artery stenting in patients with stable angina. Am J Cardiol 1998; 82:515–518.
- Casscells W, Hathorn B, David M, Krabach T, Vaughn WK, McAllister HA, Bearman G, Willerson JT. Thermal detection of cellular infiltrates in living atherosclerotic plaques: possible implications for plaque rupture and thrombosis. Lancet 1996; 347:1447–1451.
- 69. Stefanadis C, Diamantopoulos L, Vlachopoulos C, Tsiamis E, Dernellis J, Toutouzas K, Stefanadi E, Toutouzas P. Thermal heterogeneity within human atherosclerotic coronary arteries detected in vivo: a new method of detection by application of a special thermography catheter. Circulation 1999; 99:1965–1971.
- Ridker PM, Glynn RJ, Hennekens CH. C-reactive protein adds to the predictive value of total and HDL cholesterol in determining risk of first myocardial infarction. Circulation 1998; 97:2007–2011.
- Ridker PM, Buring JE, Shih J, Matias M, Hennekens CH. Prospective study of C-reactive protein and the risk of future cardiovascular events among apparently healthy women. Circulation 1998; 98:731–733.
- Koenig W, Sund M, Frohlich M, Fischer HG, Lowel H, Doring A, Hutchinson WL, Pepys MB. C-Reactive protein, a sensitive marker of inflammation, predicts future risk of coronary heart disease in initially healthy middle-aged men: results from the monica (Monitoring Trends and Determinants in Cardiovascular Disease) Augsburg Cohort Study, 1984 to 1992. Circulation 1999; 99:237–242.
- Lagrand WK, Visser CA, Hermens WT, Niessen HW, Verheugt FW, Wolbink GI, Hack CE. C-reactive protein as a cardiovascular risk factor: more than an epiphenomenon? Circulation 1999; 100:96–102.
- Toschi V, Gallo R, Lettino M, Fallon JT, Gertz SD, Fernandez-Oniz A, Chesebro JH, Badimon L, Nemerson Y, Fusler V, Badimon JJ. Tissue factor modulates the thrombogenicity of human atherosclerotic plaques. Circulation 1997; 95:594–599.
- Pepys MB, Rowe IF, Baltz ML. C-reactive protein: binding to lipids and lipoproteins. Int Rev Exp Pathol 1985; 27:83–111.
- Volanakis JE. Complement activation by C-reactive protein complexes. Ann NY Acad Sci 1982; 389:235–250.
- Cermak J, Key NS, Bach RR, Balla J, Jacob HS, Vercellotti GM. C-reactive protein induces human peripheral blood monocytes to synthesize tissue factor. Blood 1993; 82:513-520.
- Rus HG, Niculescu F, Vlaicu R. Tumor necrosis factor-alpha in human arterial wall with atherosclerosis. Atherosclerosis 1991; 89:247–254.

SII-140 CLEVELAND CLINIC JOURNAL OF MEDICINE

Downloaded from www.ccjm.org on June 3, 2025. For personal use only. All other uses require permission.

- Horrevoets AJ, Fontijn RD, van Zonneveld AJ, de Vries CJ, ten Cate JW, Pannekoek H. Vascular endothelial genes that are responsive to tumor necrosis factor-alpha in vitro are expressed in atherosclerotic lesions, including inhibitor of apoptosis protein-1, stannin, and two novel genes. Blood 1999; 93: 3418–3431.
- Ahmad M, Theofanidis P, Medford RM. Role of activating protein-1 in the regulation of the vascular cell adhesion molecule-1 gene expression by tumor necrosis factor-alpha. J Biol Chem 1998; 273:4616–4621.
- Morisaki N, Xu QP, Koshikawa T, Saito Y, Yoshida S, Ueda S. Tumour necrocis factor-alpha can modulate the phenotype of aortic smooth muscle cells. Scand J Clin Lab Invest 1993; 53:347–352.
- 82. Weber C, Draude G, Weber KS, Wubert J, Lorenz RL, Weber PC. Downregulation by tumor necrosis factor-alpha of monocyte CCR2 expression and monocyte chemotactic protein-1-induced transendothelial migration is antagonized by oxidized low-density lipoprotein: a potential mechanism of monocyte retention in atherosclerotic lesions. Atherosclerosis 1999; 145:115–123.
- Barks JL, McQuillan JJ, Iademarco MF. TNF-alpha and IL-4 synergistically increase vascular cell adhesion molecule-1 expression in cultured vascular smooth muscle cells. J Immunol 1997; 159:4532–4538.
- Libby P, Sukhova G, Lee RT, Galis ZS. Cytokines reculate vascular functions related to stability of the atherosclerotic plaque. J Cardiovasc Pharmacol 1995; 25:S9–S12.
- Galis ZS, Muszynski M, Sukhova GK, Simon-Morrissey E, Unemori EN, Lark MW, Amento E, Libby P. Cytokine-stimulated human vascular smooth muscle cells synthesize a complement of enzymes required for extracellular matrix digestion. Circ Res 1994; 75:181–189.
- Rajavashisth TB, Xu X, Jovinge S, Meisel S, Xu XO, Chai NN, Fishbein MC, Kaul S, Cercek B, Sharifi B, Shah PK. Membrane type 1 matrix metalloproteinase expression in human atherosclerotic plaques: evidence for activation by proinflammatory, mediators. Circulation 1999; 99:3103–3109.
- Dosquet C, Weill D, Wautler JL. Cytokines and thrombosis. J Cardiovasc Pharmacol 1995; 25:S13–S19.
- Charles P, Elliott MJ, Davis D, Potter A, Kalden JR, Antoni C, Breedveld FC, Smolen JS, Eberl G, deWoody K, Feldmann M, Maini RN. Regulation of cytokines, cytokine inhibitors, and acute-phase proteins following anti-TNF-alpha therapy in rheumatoid arthritis. J Immunol 1999; 163:1521–1528.
- Biasucci LM, Liuzzo G, Fantuzzi G, Caligiuri G, Rebuzzi AG, Ginnetti F, Dinarello CA, Maseri A. Increasing levels of interleukin (IL)-1Ra and IL-6 during the first 2 days of hospitalization in unstable angina are associated with increased risk of in-hospital coronary events. Circulation 1999; 99:2079–2084.
- Miyao Y, Yasue H, Ogawa H, Misumi I, Masuda T, Sakamoto T, Morita E. Elevated plasma interleukin-6 levels in patients with acute myocardial infarction. Am Heart J 1993; 126:1299–1304.
- Kushner I, Ganapathi M, Schultz D. The acute phase response is mediated by heterogeneous mechanisms. Ann NY Acad Sci 1989; 557:19–29.
- Bevilacqua MP, Schleef RR, Gimbrone MA Jr, Loskutoff DJ. Regulation of the fibrinolytic system of cultured human vascular endothelium by interleukin 1. J Clin Invest 1986; 78:587–591.
- Youker K, Smith CW, Anderson DC, Miller D, Michael LH, Rossen RD, Entman ML. Neutrophil adherence to isolated adult cardiac myocytes. Induction by cardiac lymph collected during ischemia and reperfusion. J Clin Invest 1992; 89:602–609.
- Finkel MS, Oddis C, Jacob TD, Watkins SC, Hattler BG, Simmons RL. Negative inotropic effects of cytokines on the heart mediated by nitric oxide. Science 1992; 257:387–389.
- Pannitteri G, Marino B, Campa PP, Martucci R, Testa U, Peschle C. Interleukins 6 and 8 as mediators of acute phase response in acute myocardial infarction. Am J Cardiol 1997; 80:622–625.
- RE, Garcia-Zepedia EA, Limm YC, Yoshida M, Ding HA, Gimbrone MA Jr, Luster AD, Luscinskas FW, Rosenzweig A. MCP-1 and IL-8 trigger firm adhesion of monocytes to vascular endothelium under flow conditions. Nature 1999; 398:718–723.
- Yue TL, McKenna PJ, Gu JL, Feuerstein GZ. Interleukin-8 is chemotactic for vascular smooth muscle cells. Eur J Pharmacol 1993; 240:81–84.

- Folcik VA, Aamir R, Cathcart MK. Cytokine modulation of LDL oxidation by activated human monocytes. Arterioscler Thromb Vasc Biol 1997; 17:1954–1961.
- Edgington TS, Mackman N, Brand K, Ruf W. The structural biology of expression and function of tissue factor. Thromb Haemost 1991; 66:67–79.
- 100. Nie Q, Fan J, Haraoka S, Shimokama T, Watanabe T. Inhibition of mononuclear cell recruitment in aortic intima by treatment with anti-ICAM-1 and anti-LFA-1 monoclonal antibodies in hypercholesterolemic rats: implications of the ICAM-1 and LFA-1 pathway in atherogenesis. Lab Invest 1997; 77:469–482.
- Mantovani A, Garlanda C, Introna M, Vecchi A. Regulation of endothelial cell function by pro- and anti-inflammatory cytokines. Transplant Proc 1998; 30:4239–4243.
- Baeuerle PA. I-kappaB-NF-kappaB structures: at the interface of inflammation control. Cell 1998; 95:729–731.
- Mercurio F, Manning AM. Multiple signals converging on NFkappaB. Curr Opin Cell Biol 1999; 11:226–232.
- Lee JI, Burckart GJ. Nuclear factor kappa B: important transcription factor and therapeutic target. J Clin Pharmacol 1998; 38:981–993.
- Barnes PJ, Karin M. Nuclear factor-kappaB: a pivotal transcription factor in chronic inflammatory diseases. N Engl J Med 1997; 336:1066–1071.
- Massi-Benedetti M, Federici MO. Cardiovascular risk factors in type 2 diabetes: the role of hyperglycaemia. Exp Clin Endocrinol Diabetes 1999; 107:S120–S123.
- Dichtl W, Nilsson L, Goncalves I, Ares MP, Banfi C, Calara F, Hamsten A, Eriksson P, Nilsson J. Very low-density lipoprotein activates nuclear factor-kappaB in endothelial cells. Circ Res 1999; 84:1085–1094.
- 108. Brand K, Eisele T, Kreusel U, Page M, Page S, Haas M, Gerling A, Kaltschmidt C, Neumann FJ, Mackman N, Baeurele PA, Walli AK, Neumeier D. Dysregulation of monocytic nuclear factorkappa B by oxidized low-density lipoprotein. Arterioscler Thromb Vasc Biol 1997; 17:1901–1909.
- Kranzhofer R, Browatzki M, Schmidt J, Kubler W. Angiotensin II activates the proinflammatory transcription factor nuclear factorkappaB in human monocytes. Biochem Biophys Res Commun 1999; 257:826–828.
- Yerneni KK, Bai W, Khan BV, Medford RM, Natarajan R. Hyperglycemia-induced activation of nuclear transcription factor kappaB in vascular smooth muscle cells. Diabetes 1999; 48:855–864.
- 111. Bustos C, Hernandez-Presa MA, Ortego M, Tunon J, Ortega L, Perez F, Diaz C, Hernandez G, Egido J. HMG-CoA reductase inhibition by atorvastatin reduces neointimal inflammation in a rabbit model of atherosclerosis. J Am Coll Cardiol 1998; 32:2057–2064.
- Ritchie ME. Nuclear factor-kappaB is selectively and markedly activated in humans with unstable angina pectoris. Circulation 1998; 98:1707–1713.
- 113. Kopp E, Ghosh S. Inhibition of NF-kappa B by sodium salicylate and aspirin. Science 1994; 265:956–959.
- 114. Weber C, Erl W, Pietsch A, Weber PC. Aspirin inhibits nuclear factor-kappa B mobilization and monocyte adhesion in stimulated human endothelial cells. Circulation 1995; 91:1914–1917.
- Mach F, Schonbeck U, Libby P. CD40 signaling in vascular cells: a key role in atherosclerosis? Atherosclerosis 1998; 137(suppl):S89–S95.
- 116. Mohan C, Shi Y, Laman JD, Datta SK. Interaction between CD40 and its ligand gp39 in the development of murine lupus nephritis. J Immunol 1995; 154:1470–1480.
- 117. Larsen CP, Elwood ET, Alexander DZ, Ritchie SC, Hendrix R, Tucker-Burden C, Cho HR, Aruffo A, Hollenbaugh D, Linsley PS, Winn KJ, Pearson TC. Long-term acceptance of skin and cardiac allografts after blocking CD40 and CD28 pathways. Nature 1996; 381:434–438.
- Gerritse K, Laman JD, Noelle RJ, Aruffo A, Ledbetter JA, Boersma WJ, Claassen E. CD40-CD40 ligand interactions in experimental allergic encephalomyelitis and multiple sclerosis. Proc Natl Acad Sci USA 1996; 93:2499–2504.
- Mach F, Schonbeck U, Sukhova GK, Atkinson E, Libby P. Reduction of atherosclerosis in mice by inhibition of CD40 signalling. Nature 1998; 394:200–203.

VOLUME 69 • SUPPLEMENT II

CLEVELAND CLINIC JOURNAL OF MEDICINE SII-141

- 120. Aukrust P, Muller F, Ueland T, Berget T, Aaser, Brunsvig A, Solum NO, Forfang K, Froland SS, Gullestad L . Enhanced levels of soluble and membrane-bound CD40 ligand in patients with unstable angina: possible reflection of T lymphocyte and platelet involvement in the pathogenesis of acute coronary syndromes. Circulation 1999; 100:614–620.
- 121. Plutzky J. Atherosclerotic plaque rupture: emerging insights and opportunities. Am J Cardiol 1999; 84:15J–20J.
- Ricote M, Li AC, Willson TM, Kelly CJ, Glass CK. The peroxisome proliferator-activated receptor-gamma is a negative regulator of macrophage activation. Nature 1998; 391:79–82.
- 123. Nagy L, Tontonoz P, Alvarez JG, Chen H, Evans RM. Oxidized LDL regulates macrophage gene expression through ligand activation of PPARgamma. Cell 1998; 93:229–240.
- 124. Tontonoz P, Nagy L, Alvarez JG, Thomazy VA, Evans RM. PPARgamma promotes monocyte/macrophage differentiation and uptake of oxidized LDL. Cell 1998; 93:241–252.
- 125. Marx N, Sukhova G, Murphy C, Libby P, Plutzky J. Macrophages in human atheroma contain PPARgamma: differentiation-dependent peroxisomal proliferator-activated receptor gamma (PPARgamma) expression and reduction of MMP-9 activity through PPARgamma activation in mononuclear phagocytes in vitro Am J Pathol 1998; 133:17–23.
- Chinetti G, Griglio S, Antonucci M, Torra IP, Delerive P, Majd Z, Fruchart JC, Chapman J, Najib J, Staels B. Activation of proliferator-activated receptors alpha and gamma induces apoptosis of human monocyte-derived macrophages. J Biol Chem 1998; 273:25573–25580.
- 127. Jackson SM, Parhami F, Xi XP, Berliner JA, Hsueh WA, Law RE, Demer LL. Peroxisome proliferator-activated receptor activators target human endothelial cells to inhibit leukocyte-endothelial cell interaction. Arterioscler Thromb Vasc Biol 1999; 19:2094-2104.
- Marx N, Sukhova GK, Collins T, Libby P, Plutzky J. PPARalpha activators inhibit cytokine-induced vascular cell adhesion molecule-1 expression in human endothelial cells. Circulation 1999; 99:3125–3131.
- 129. Delerive P, Martin-Nizard F, Chinetti G, Trottein F, Fruchart JC, Najib J, Duriez P, Staels B. Peroxisome proliferator-activated receptor activators inhibit thrombin-induced endothelin-1 production in human vascular endothelia1 cells by inhibiting the activator protein-1 signaling pathway. Circ Res 1999; 85:394–402.
- 130. Staels B, Koenig W, Habib A, Merval R, Lebret M, Torra IP, Delerive P, Fadel A, Chinetti G, Fruchart JC, Najib J, Maclouf J, Tedgui A. Activation of human aortic smooth-muscle cells is inhibited by PPARalpha but not by PPARgamma activators. Nature 1998; 393:790–793.
- 131. Rubins HB, Robins SJ, Collins D, Fye CL, Anderson JW, Elam MB, Faas FH, Linares E, Schaefer EJ, Schectman G, Wilt TJ, Wittes J. Gemfibrozil for the secondary prevention of coronary heart disease in men with low levels of high-density lipoprotein cholesterol. Veterans Affairs High-Density Lipoprotein Cholesterol Intervention Trial Study Group. N Engl J Med 1999; 341:410–418.
- Dubois RN, Abramson SB, Crofford L, Gupta RA, Simon LS, Van de Putte LB, Lipsky PE. Cyclooxygenase in biology and disease. FASEB J 1998; 12:1063–1073.
- Cryer B, Dubois A. The advent of highly selective inhibitors of cyclooxygenase—a review. Prostaglandins Other Lipid Mediat 1998; 56:341–361.
- 134. Xie WL, Chipman JG, Robertson DL, Erikson RL, Simmons DL. Expression of a mitogen-responsive gene encoding prostaglandin synthase is regulated by mRNA splicing. Proc Natl Acad Sci USA 1991; 88:2692–2696.
- 135. Seibert K, Masferrer J, Zhang Y, Gregory S, Olson G, Hauser S, Leahy K, Perkins W, Isakson P. Mediation of inflammation by cyclooxygenase-2. Agents Actions Suppl 1995; 46:41–50.

- 136. Baker CS, Hall RJ, Evans TJ, Pomerance A, Maclouf J, Creminon C, Yacoub MH, Polak JM. Cyclooxygenase-2 is widely expressed in atherosclerotic lesions affecting native and transplanted human coronary arteries and colocalizes with inducible nitric oxide synthase and nitrotyrosine particularly in macrophages. Arterioscler Thromb Vasc Biol 1999; 19:646–655.
- 137. Schonbeck U, Sukhova GK, Graber P, Coulter S, Libby P. Augmented expression of cyclooxygenase-2 in human atherosclerotic lesions. Am J Pathol 1999; 155:1281–1291.
- 138. Niiro H, Otsuka T, Tanabe T, Hara S, Kuga S, Nemoto Y, Tanaka Y, Nakashima H, Kitajima S, Abe M. Inhibition by interleukin-10 of inducible cyclooxygenase expression in lipopolysaccharide-stimulated monocytes: its underlying mechanism in comparison with interleukin-4. Blood 1995; 85:3736–3745.
- Arias-Negrete S, Keller K, Chadee K. Proinflammatory cytokines regulate cyclooxygenase-2 mRNA expression in human macrophages. Biochem Biophys Res Commun 1995; 208:582–589.
- 140. Speir E, Yu ZX, Ferrans VJ, Huang ES, Epstein SE. Aspirin attenuates cytomegalovirus infectivity and gene expression mediated by cyclooxygenase-2 in coronary artery smooth muscle cells. Circ Res 1998; 83:210–216.
- Cockerill GW, Rye KA, Gamble JR, Vadas MA, Barter PJ. Highdensity lipoproteins inhibit cytokine-induced expression of endothelial cell adhesion molecules. Arterioscler Thromb Vasc Biol 1995; 15:1987–1994.
- 142. Bishop-Bailey D, Burke-Gaffney A, Hellewell PG, Pepper JR, Mitchell JA. Cyclo-oxygenase-2 regulates inducible ICAM-1 and VCAM-1 expression in human vascular smooth muscle cells. Biochem Biophys Res Commun 1998; 249:44–47.
- 143. Sajadieh A, Wendelboe O, Hansen JF, Mortensen LS. Nonsteroidal anti-inflammatory drugs after acute myocardial infarction. DAVIT Study Group. Danish Verapamil Infarction Trial. Am J Cardiol 1999; 83:1263–1265, A9.
- 144. Ridker PM, Cushman M, Stampfer MJ, Tracy RP, Hennekens CH. Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men [published erratum appears in N Engl J Med 1997;337(5):356]. N Engl J Med 1997; 336:973–979.
- 145. Sacks FM, Ridker PM. Lipid lowering and beyond: results from the CARE study on lipoproteins and inflammation. Cholesterol and Recurrent Events. Herz 1999; 24:51–56.
- 146. Ridker PM. Rifai N, Pfeffer MA, Sacks FM, Moye LA, Goldman S, Flaker GC, Braunwald E. Inflammation, pravastatin, and the risk of coronary events after myocardial infarction in patients with average cholesterol levels. Cholesterol and Recurrent Events (CARE) Investigators. Circulation 1998; 98: 839-844.
- Vaughan CJ, Murphy MB, Buckley BM. Statins do more than just lower cholesterol. Lancet 1996; 348:1079–1082.
- McPherson R, Tsoukas C, Baines MG, Vost A, Melino MR, Zupkis RV, Pross HF. Effects of lovastatin on natural killer cell function and other immunological parameters in man. J Clin Immunol 1993; 13:439–444.
- 149. Bellosta S, Via D, Canavesi M, Pfister P, Fumagalli R, Paoletti R, Bernini F. HMG-CoA reductase inhibitors reduce MMP-9 secretion by macrophages. Arterioscler Thromb Vasc Biol 1998; 18:1671–1678.
- Cheng JW, Ngo MN. Current perspective on the use of angiotensin-converting enzyme inhibitors in the management of coronary (atherosclerotic) artery disease. Ann Pharmacother 1997; 31:1499-1506.
- 151. HOPE Investigators. Effects of an angiotensin -converting-enzyme inhibitor, ramapril, on death from cardiovascular causes, myocardial infarction, stroke in high-risk patients. N Engl J Med 2000. In press.
- Faxon DP GR, Chronos NA, Gurbel PA, Martin JS. The effect of CD11/CD18 inhibitor (Hu23F2G) on infarct size following direct angioplasty: the HALT MI study. Circulation 1999; 100:I-791.
- 153. Pasceri et al. Circulation 1999; 100:2124-2126.