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# Hepatitis C virus: Prevention, screening, and interpretation of assays

## ABSTRACT

Patients at risk of hepatitis C virus (HCV) infection should be screened for it so that they can be treated and potentially cured, or can at least avoid transmitting the disease to others. The authors describe why and how to screen for HCV and how to interpret the test results.

## **KEY POINTS**

Patients who should be screened include intravenous drug abusers, people infected with human immunodeficiency virus, patients with unexplained elevated alanine aminotransferase levels, infants born to infected mothers, and people with infected sexual partners.

Patients at risk of HCV infection should be tested for anti-HCV antibody using an enzyme immunoassay (EIA).

Positive results on anti-HCV EIA testing should be confirmed with an assay for HCV RNA.

HCV genotyping can help predict the response to therapy. Genotypes 2 or 3 are more likely to respond to therapy than genotype 1.

**S** CREENING FOR HEPATITIS C VIRUS (HCV) infection in high-risk populations can identify, early on, people at risk of progressive liver disease who may benefit from antiviral therapy and counseling. The US Centers for Disease Control and Prevention (CDC) recommends that all people be assessed for HCV risk factors and that those with risk factors be screened for HCV antibodies (anti-HCV),<sup>1</sup> and members of the national societies of gastroenterology and hepatology have endorsed this recommendation.<sup>2</sup>

Unfortunately, rates at which primary care patients are assessed for risk factors and the rates at which patients at higher risk are screened remain below the goals set by the CDC.<sup>3-6</sup> All health care practitioners need to understand how to establish or exclude a diagnosis of HCV infection and to interpret the tests correctly.

## WHY SCREEN FOR HCV?

HCV infection is a major public health problem and a leading cause of chronic liver disease. In the United States, an estimated 3.2 million persons (1.3% of the population) have been infected.<sup>7</sup> However, in the inner-city primary care setting the rate of HCV infection is as high as 8%, and in Veterans Administration populations it is 17%.<sup>8,9</sup> The worldwide prevalence of HCV infection is 2.0%, corresponding to 140 million persons.

Screening of blood products has led to a decline in the incidence of acute hepatitis C since the late 1980s, although rates have reached a plateau in recent years (**FIGURE 1**).<sup>10</sup>

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Approximately 20% of patients infected with HCV develop a serious sequela, such as severe fibrosis, cirrhosis, end-stage liver disease, or hepatocellular carcinoma. Currently, HCV infection causes an estimated 8,000 to 10,000 deaths annually in the United States, and that number is predicted to triple in the next 10 to 20 years. Furthermore, HCV-related disease is the leading indication for liver transplantation in the United States, and it is estimated to cost \$600 million to \$1 billion annually in medical expenses and loss of work.8

#### Screening can reduce adverse outcomes

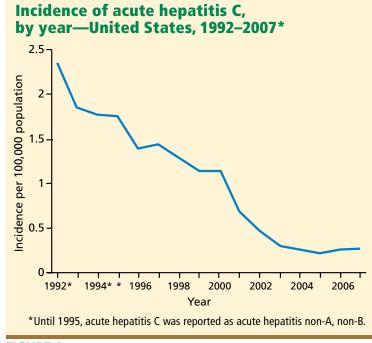
HCV screening has several potential benefits. By detecting HCV infection early, screening facilitates virologic suppression, as treatment earlier in the course of the disease is more effective than later.<sup>11,12</sup> Further, early diagnosis together with patient education and subsequent lifestyle modifications may reduce the risk of transmission of HCV infection to other people.13,14

Antiviral therapy with pegylated interferons and ribavirin can cure hepatitis C in up to 90% of cases, depending on the viral genotype<sup>15-17</sup> (see discussion of HCV genotypes below). In addition, treatment slows the progression of fibrosis.<sup>18</sup> The incidence of hepatocellular carcinoma is lower in patients who achieve a sustained virologic response to antiviral therapy.<sup>19</sup> Finally, antiviral therapy prolongs survival.<sup>20</sup>

New drug therapies are being developed and may, we hope, be even more effective than current drugs. Inhibitors of HCV-specific enzymes such as NS3/4 protease, combined with pegylated interferons and ribavirin, are in phase III clinical trials. These drugs are expected to be available for clinical practice within the next 2 years.<sup>21–23</sup> Additionally, nitazoxanide (Alinia), an inducer of eIF2a and PKR phosphorylation, has been shown to increase the treatment response to HCV genotype 4. Studies<sup>24</sup> are currently under way in patients infected with HCV genotype 1.

### Screening is cost-effective

The National Hepatitis Surveillance Program<sup>25</sup> calculated the cost of screening for HCV to be \$1,246 per case detected. How-



#### **FIGURE 1**

DANIELS D, GRYTDAL S, WASLEY A; US CENTERS FOR DISEASE CONTROL AND PREVENTION. SURVEILLANCE FOR ACUTE VIRAL HEPATITIS-UNITED STATES, 2007. MMWR SURVEILL SUMM 2009; 58:SS-3.

ever, a more vigorous analysis of the same data using several different models to incorporate risk factors based on history revealed drug abuse costs between \$357 and \$1,047 per case detected. This compares favorably with the cost of screening for other diseases that physicians risk factor for routinely screen for.

Antiviral combination therapy for chronic hepatitis C has been shown to be effective in terms of quality-adjusted life-years gained and cost-effectiveness in several studies.<sup>26–28</sup>

### HOW TO SCREEN

The optimal approach to screening for HCV is to look for a history of risk of exposure to the virus and then to test those who have risk factors (TABLE 1).

To test everyone in the general population would be neither cost-effective nor practical, which is why the CDC recommends that serologic screening for HCV infection be done only in people who have well-established risk factors for it.<sup>1,5</sup>

Therefore, screening should begin by obtaining a relevant medical history as part of

Intravenous is the strongest **HCV** infection

## TABLE 1

## **Risk factors for hepatitis C virus (HCV) infection**

Injection drug use (past or present)

Receipt of clotting factor concentrates before 1987

Receipt of blood or blood components (red cells, platelets, fresh-frozen plasma)

Receipt of blood from an HCV-positive donor

Repeatedly elevated (or unexplained intermittently elevated) serum alanine aminotransferase levels

Long-term hemodialysis

Specific high-risk exposure to known HCV-positive blood in health care workers (needlestick, mucosal exposure via "splash accident")

Human immunodeficiency virus infection

Being the child of an HCV-positive woman

History of multiple sex partners or sexually transmitted infections

a routine health evaluation. But how should this be done?

McGinn et al<sup>29</sup> asked 1,000 patients attending an inner-city clinic to fill out a 27item questionnaire assessing five "domains" of risk factors for HCV: work, medical, exposure, personal care, and social history. Afterward, they tested all 1,000 patients. They found that the risk factors that best predicted positive results on testing were in three domains: medical (eg, blood transfusions, dialysis, other medical procedures, and elevated liver enzymes), exposure (past contact with another person's blood), and social history (eg, illicit drug use, incarceration, and sexual activity).

The National Hepatitis Surveillance Program<sup>25</sup> explored the cost and yield of several screening strategies for hepatitis C, ie, testing only in patients who had a greater than 7% likelihood of infection based on an empirically derived mathematical model; testing only if significant risk factors were revealed in a simple questionnaire; or testing only if the alanine aminotransferase (ALT) level was elevated. The predictive mathematical model was the most effective and efficient means of deciding who should be tested.

Unfortunately, such a model is too cumbersome to be clinically applicable, and clinical prediction tools for HCV screening have been underused.

## GROUPS AT HIGH RISK OF HCV

Groups at risk of HCV infection can be classified as being at high, intermediate, or low risk. The American Association for the Study of Liver Diseases<sup>2</sup> rates the level of evidence for screening in all of the following risk groups as class I (ie, there is evidence or general agreement that it is beneficial, useful, and effective) and level B (ie, the data are derived from nonrandomized studies).

### Intravenous drug abusers

Intravenous drug abuse is the strongest independent risk factor for HCV infection.<sup>30–33</sup> It has been the main route of HCV infection over the past decades and currently accounts for 60% of HCV transmission in the United States.<sup>7,10,34–37</sup>

### Hemophilia patients treated with clotting factor concentrates produced before 1987

HCV seroprevalence is very high in patients with hemophilia who received infusions of plasma-derived clotting factor concentrates before 1987.<sup>38</sup> In these patients, the HCV genotypes are predominantly 1 and 3, and to a lesser extent genotype 2.<sup>39,40</sup> These genotypes likely reflect the prior exposures of the plasma donors.<sup>41</sup> (See discussion of HCV genotypes below.) Individuals receiv-

If ALT or AST is elevated, first repeat the test

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ing clotting factor concentrates prepared from plasma pools were at high risk of HCV infection until effective procedures to inactivate viruses were introduced in 1985 (factor VIII) and 1987 (factor IX).42

## People infected with HIV

About 25% of people infected with human immunodeficiency virus (HIV) in the Western world also have chronic HCV infection.<sup>43</sup> Progression of liver disease is accelerated in HIV-HCV coinfection, and the risk of cirrhosis is twice as high.<sup>44</sup>

However, about 6% of HIV-positive patients fail to develop HCV antibodies when infected. Thus, HCV RNA should be assessed in HIV patients with unexplained liver disease who are negative for anti-HCV.45

The distribution of HCV genotypes in HIV-infected patients reflects the route of transmission. Genotype 1b accounts for 66% of posttransfusion HCV infections, while genotypes 1a and 3a are more common in intravenous drug users.

## **GROUPS AT INTERMEDIATE RISK OF HCV**

## **Recipients of blood transfusions** before 1992

Before 1992, blood transfusions carried a risk of HCV infection of up to 7% with each unit transfused. Prospective studies of transfusion recipients in the United States found that rates of posttransfusion hepatitis in the 1960s exceeded 20%,<sup>36</sup> since most patients received multiple units of blood.

In the mid-1970s, before HCV had been identified, available diagnostic tests indicated that 90% of cases of posttransfusion hepatitis were not caused by hepatitis A or hepatitis B viruses. By this time, the move to all-volunteer blood donors instead of paid donors had reduced the risk of posttransfusion hepatitis to 10%, 22, 37, 46

Although non-A, non-B hepatitis was first recognized because of its association with blood transfusion, population-based sentinel surveillance showed that it accounted for 15% to 20% of cases of community-acquired viral hepatitis in the United States.<sup>35</sup> The advent of molecular cloning in 1988 indicated that

non-A, non-B hepatitis was primarily caused by HCV.47-52

Screening of blood has reduced the rate of posttransfusion hepatitis C by a factor of about 10,000, to a current rate of 1 per million transfusions.53 The few cases that still occur are due to newly infected people donating blood before they have developed antibodies to the virus, which can take up to 8 weeks.54

## Recipients of solid-organ transplants before 1992

Before organ donors were screened for HCV, recipients of solid-organ transplants from infected donors had a high risk of acquiring HCV infection. Transmission rates in different cohorts ranged from 30% to 80%.55 In an attempt to improve the safety of organ transplantation, many transplant centers now screen donors for anti-HCV and test for HCV RNA for verification.

A related problem is pre-existing HCV infection in transplant recipients. Izopet et al<sup>56</sup> reported that, in renal transplant recipients with preexisting HCV infection, the HCV RNA titer rose about 10 times (1 log) higher after transplantation, owing to the immunosuppressive drugs that transplant recipients must take. Although this higher viral load does not affect the progression of fibrosis in all patients, the effect of immunosuppressive sooner than therapy on liver disease results in a worse outcome for some, and it reduces survival beginning in the second decade after kidney transplantation.56

Additionally, treatment of HCV infection in transplant recipients may pose a challenge, as those receiving immunosuppressive therapy with tacrolimus (Prograf) or cyclosporine (Sandimmune) may develop some degree of renal insufficiency, complicating the use of ribavirin (Rebetol) and subjecting patients to a higher risk of severe anemia. Furthermore, interferon therapy increases the risk of renal allograft rejection and, accordingly, is not often used in renal transplant recipients.

## Patients with unexplained elevated aminotransferase levels

HCV infection affects an estimated 1.8% of the general population, but the rate is much

**Screen children** of HCV-positive mothers no 12 months

higher in people with ALT levels over 40 U/L. Most patients with chronic hepatitis C have no symptoms or only mild symptoms and minimally elevated levels of ALT and aspartate aminotransferase (AST)—ie, two to five times higher than the upper limit of normal.

The first step in the workup of aminotransferase elevations is to confirm the abnormality by repeating the blood test. If an elevation is confirmed, further investigation is warranted. A directed history and physical examination is important and may disclose risk factors, raising clinical suspicion of a particular disease.

Some caveats: The proportion of patients with HCV viremia who have abnormally high aminotransferase levels ranges between only 54% and 66%.<sup>57–59</sup> In patients with risk factors for HCV infection and abnormal liver enzyme levels, HCV infection is probable but not certain. Also, liver enzyme tests do not reveal the extent of hepatic injury or reflect the true status of hepatic function.<sup>60</sup>

## Infants born to infected mothers

Children born to HCV-positive women should be tested for anti-HCV no sooner than age 12 months, when passively transferred maternal anti-HCV declines below detectable levels. If earlier diagnosis of HCV infection is desired, a real-time polymerase chain reaction (PCR) test for HCV RNA can be done at or after the infant's first "well-child" visit at age 1 to 2 months.

If positive for either anti-HCV or HCV RNA, children should be evaluated for liver disease, and those with persistently elevated ALT levels should be referred to a specialist for medical management.<sup>2,5</sup>

## GROUPS AT LOW RISK OF HCV

## People who have had sexual relations with multiple or infected partners

Sexual activity is associated with a low but measurable risk of transmission of HCV. Large population-based studies, including the National Hepatitis Surveillance Program,<sup>25</sup> found an independent association between HCV infection and having sexual relations with multiple partners or with a partner who is infected with HCV.

The CDC reported that 15% to 20% of patients with acute hepatitis C had a history of sexual exposure but no other risk factors. Two-thirds of them had an anti-HCV-positive sexual partner, and one-third reported having had more than two partners in the 6 months before illness.<sup>5</sup>

More data are needed to determine the risk of and the factors related to transmission of HCV between long-term steady partners as well as in persons with high-risk sexual practices, including whether other sexually transmitted diseases promote transmission of HCV by influencing viral load or modifying mucosal barriers.

## Health care workers exposed to HCV, eg, by needlestick

The prevalence of HCV infection in health care workers is no greater than that in the general population, averaging 1% to 2%, and is actually 10 times lower than that of hepatitis B virus infection.<sup>47,48,61,62</sup>

However, within the disciplines, some groups have a higher prevalence of HCV infection, suggesting that some occupations carry a higher risk. In two US studies, the prevalence of HCV infection was higher in oral surgeons (2.0% and 9.3%) than in other dentists (0.7% and 0.97%).<sup>63,64</sup>

In a single study that evaluated risk factors for infection, a history of needlestick injury was the only occupational risk factor that was independently associated with HCV infection.<sup>65</sup> The average incidence of anti-HCV seroconversion after a needlestick or after an injury with a sharp object contaminated by an HCV-positive source is 1.8% (range 0%–7%).<sup>66–69</sup>

Although no studies of incidence have documented transmission via mucous membrane or nonintact skin exposures, transmission of HCV from blood splashes to the conjunctiva have been described.<sup>70,71</sup>

Refer to TABLE 2 for postexposure follow-up recommendations.

It is worth noting that exposure to blood from unclean needles used in tattooing or body piercing also confers a risk of HCV infection.

Incidence of seroconversion after accidental stick with a needle contaminated with HCV: 1.8%

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## SEROLOGIC SCREENING TESTS FOR HCV

FIGURE 2 is an algorithm for laboratory investigation of suspected HCV infection,<sup>72</sup> TABLE 3 summarizes how to interpret the test results, and TABLE 4 lists how the various tests are used in diagnosing HCV infection, estimating the prognosis, and treating HCV infection.<sup>73</sup>

Two classes of assays are used to diagnose HCV infection:

- Serologic assays that detect specific antibody to HCV (anti-HCV)
- Molecular assays that detect viral RNA.

## Initial serologic screening tests for anti-HCV

Enzyme immunoassays (EIAs) are reproducible, inexpensive, and approved by the US Food and Drug Administration for diagnosing HCV infection. They are suitable for screening populations at risk and are recommended as the initial serologic test for patients with clinical liver disease.

Two EIAs are approved for clinical use:

- Abbott HCV EIA 2.0 (Abbott Laboratories, Abbott Park, IL)
- Ortho HCV Version 3.0 enzyme-linked immunosorbent assay (ELISA) (Ortho-Clinical Diagnostics, Rochester, NY).

One enhanced chemiluminescence immunoassay is also approved:

• Vitros Anti-HCV assay (Ortho-Clinical Diagnostics). In practical terms, this test is equivalent to the two EIAs, and the discussion below about EIAs applies to this test as well.

These third-generation tests are highly sensitive (> 99%) and specific (99%) in immunocompetent patients, and eliminate the need for a confirmatory immunoblot assay in patients with clinical liver disease, particularly those with risk factors for HCV infection.

False-positive results are rare now, but they were common with earlier generations of these assays. Most false-positive results occur in patients with autoimmune liver disease or hypergammaglobulinemia who have normal liver enzyme levels and no risk factors for HCV infection. In fact, *all* positive anti-HCV results should be followed up with an HCV RNA test.

False-negative results are also uncommon, usually occurring only in immunosuppressed

## TABLE 2

## Follow-up of workers potentially exposed to hepatitis C virus

#### For the source:

Baseline testing for antibodies to hepatitis C virus (anti-HCV)

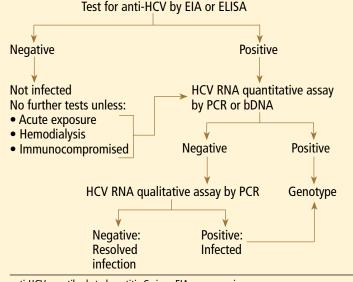
#### For the worker exposed to an HCV-positive source: Baseline testing, including anti-HCV antibodies and alanine aminotranferase (ALT)

Follow-up testing for anti-HCV and ALT at 4–6 months (if earlier diagnosis of HCV infection is desired, testing for HCV RNA may be performed at 4–6 weeks)

#### Confirmation by supplemental anti-HCV testing of all anti-HCV results reported as positive by enzyme immunoassay

ADAPTED FROM US PUBLIC HEALTH SERVICE. UPDATED US PUBLIC HEALTH SERVICE GUIDELINES FOR THE MANAGEMENT OF OCCUPATIONAL EXPOSURES TO HBV, HCV, AND HIV AND RECOM-MENDATIONS FOR POSTEXPOSURE PROPHYLAXIS. MMWR RECOMM REP 2001; 50(RR-11):1–52.

## Algorithm for laboratory investigation of suspected hepatitis C virus infection



anti-HCV = antibody to hepatitis C virus; EIA = enzyme immune assay; ELISA = enzyme-linked immunosorbent assay; PCR = polymerase chain reaction; bDNA = branched DNA

ADAPTED FROM CAREY W. TESTS AND SCREENING STRATEGIES FOR THE DIAGNOSIS OF HEPATITIS C. CLEVE CLIN J MED 2003; 70(SUPPL 4):S7–S13.

## FIGURE 2

patients (eg, organ transplant recipients and HIV-positive patients) and in patients on long-term hemodialysis. Therefore, patients with a history of hemodialysis should be considered for an HCV RNA assay rather than an

## TABLE 3

## How to interpret hepatitis C test results

	ANTI-HCV	HCV RNA (PCR)	INTERPRETATION
	Negative	Negative	No infection
	Positive	Positive	HCV present (acute or chronic infection)
	Negative	Positive	Chronic infection in immunosuppressed patient Early infection
	Positive	Negative	Resolved infection Treated infection, HCV below detectable levels (verify with qualitative HCV RNA PCR) False-positive anti-HCV test (< 1%)

HCV = hepatitis C virus, anti-HCV = antibodies to HCV, RNA = ribonucleic acid, PCR = polymerase chain reaction

## TABLE 4 Uses of specific hepatitis C tests

USES	TESTS	
To detect acute HCV infection	HCV RNA	
To detect chronic HCV infection	EIA; if EIA is positive then HCV RNA	
To detect vertical transmission	HCV RNA	
To detect occupational exposure	HCV RNA, EIA	
To estimate prognosis	None	
To decide whether to treat	EIA, HCV RNA, HCV genotyping	
To evaluate response to treatment	HCV RNA by sensitive assay	
To test for sustained eradication	HCV RNA by sensitive assay	

HCV = hepatitis C virus, RNA = ribonucleic acid, EIA = enzyme immunoassay

ADAPTED FROM CAREY WD, JEFFERS L, KUGELMAS M, ET AL. HEPATITIS C MANAGEMENT. HEPATITIS C MONOGRAPH. CLEVELAND CLINIC; HTTP://WWW.CLEVELANDCLINICMEDED.COM/ONLINE/MONOGRAPH/HEPC/PAGE1.HTP.

> EIA. Measurement of ALT will not be useful because ALT levels are lower in patients with end-stage renal disease. In most other clinical situations, the HCV EIA is an outstanding screening test for HCV infection because of its high sensitivity and relatively low cost (< \$50).

> Although the specificity of these tests is good, the predictive value of a positive result

varies substantially by the pretest probability of HCV infection. For example, in a group of injection-drug users who are very likely to have ongoing or remote infection, all positive HCV EIA results are likely truly positive.<sup>74</sup> On the other hand, in healthy blood donors, up to half of all positive third-generation EIA tests are falsely positive.<sup>75</sup>

### Important points

- A positive anti-HCV antibody test does not distinguish acute from chronic disease or active from past infection, nor is it a sign of immunity or protection.
- A positive anti-HCV EIA requires HCV RNA measurement to discriminate between current infection on the one hand, and either resolved HCV infection or a false-positive result on the other.
- A positive EIA anti-HCV test is a marker that hepatitis C may be present, and it must be followed by confirmatory HCV RNA testing.
- Physicians should be mindful of the potential tribulations associated with falsepositive tests. A false-positive test may result in harm to patients that is difficult to measure, such as anxiety, labeling in the medical record, and detrimental effects on close relationships.

## CONFIRMATORY TESTING WITH ASSAYS FOR HCV RNA

As stated above, a positive result on an anti-HCV EIA needs to be confirmed with an assay for HCV RNA, of which there are two types, ie, qualitative and quantitative.

Each involves trade-offs. Qualitative assays are more sensitive and detect more cases, but they provide no information about the amount of virus (viral load). Quantitative assays are less sensitive, so a negative result does not completely exclude hepatitis C, although they can still can detect 95% of cases. They do, however, measure the viral load.

Therefore, the type of test to use depends on the patient's risk profile, the goals of testing, and the setting in which future care will be provided. The primary objective when a patient has a positive EIA test is to determine whether he or she has ongoing infection, a goal most expeditiously achieved using a qualitative assay. However, since a quantitative assay can detect the vast majority of cases of active HCV infection, many clinicians select this as the test of first choice when the probability of HCV is high (eg, in a patient with risk factors and abnormal liver tests). If the pretest probability is low, a qualitative assay is the better choice.

Many commercial assays are available for detecting (qualitative assays) or measuring (quantitative assays) HCV RNA.

## Qualitative HCV RNA assays

The approved qualitative assays are:

- Amplicor HCV Test, version 2.0 (Roche Molecular Diagnostics, Pleasanton, CA)
- Cobas Amplicor HCV Test, version 2.0 (Roche Molecular Diagnostics)
- Ampliscreen (Roche Molecular Diagnostics)
- Versant HCV RNA Qualitative Assay (Siemens Healthcare Diagnostics, Deerfield, IL)
- Procleix HIV-1/HCV Assay (Chiron, Emeryville, CA).

## Quantitative HCV RNA assays

The approved quantitative assays are:

- Amplicor HCV Monitor (Roche Molecular Diagnostics)
- Cobas Amplicor HCV Monitor, version 2.0 (Roche Molecular Diagnostics)
- Versant HCV RNA 3.0 Assay (bDNA) (Siemens Healthcare Diagnostics)
- Cobas Taqman HCV Test (Roche Molecular Diagnostics).

Quantitative tests use target amplification with PCR, transcription-mediated amplification (TMA), or a signal amplification technique such as a branched DNA (bDNA) assay. The sensitivity varies for different types of amplification. TMA assays appear to be the most sensitive for detecting HCV RNA.

The latest innovation is real-time PCR, which shortens the typical time for PCR processing from 1.5 hours to 35 minutes. It may also detect relapsed HCV infection earlier than regular PCR. With the recent availability of real-time PCR assays, which have sensitivities of 10 to 50 IU/mL, many experts feel there is no longer a need for qualitative assays.<sup>74</sup> In

fact, many laboratories no longer offer qualitative testing. The Cleveland Clinic laboratory has recently stopped offering this test.

Because RNA testing is widely available, the recombinant immunoblot assay (RIBA) has become obsolete in diagnosing HCV infection, except in special circumstances. Currently, the primary purpose of RIBA testing is to distinguish between resolved HCV infection (EIA-positive, HCV RNA-negative, RIBA-positive) and a false-positive EIA (EIA-positive, HCV RNA-negative, RIBAnegative).

In summary, patients suspected of having acute or chronic HCV infection should first be tested for anti-HCV. Subsequently, HCV RNA testing should be performed in:

- Patients with a positive anti-HCV test
- Patients for whom antiviral treatment is being considered (using a sensitive quantitative assay)
- Patients with unexplained liver disease whose anti-HCV test is negative and who are immunocompromised or suspected of having acute HCV infection.

## Significance of the HCV viral load

The significance of the HCV viral load is In healthy widely misunderstood. The amount of virus in **blood donors**, the blood does not correlate with symptoms, histologic liver injury, or the stage or aggressiveness of disease. Its sole importance is in of positive relation to therapy.

The HCV viral load, measured before treatment, helps predict the likelihood of a treat- are falsely ment response: the lower the pretreatment viral load, the more likely that the patient will respond to current HCV therapies.

Additionally, the pretreatment viral load serves as a baseline for comparison with subsequent measurements during treatment. Patients with HCV genotype 1 who do not achieve more than a 2-log (99%) reduction in viral load by the 12th week of treatment (an early virologic response) have a low response rate, and treatment should generally be stopped, given its cost and side effects.<sup>76</sup> However, measuring the viral load to detect an early virologic response is less helpful in patients with HCV genotype 2 or 3 infection, since these patients require only 24 weeks of therapy and most of them clear the virus by

up to half anti-HCV tests positive

week 12 and respond to therapy.

Additionally, patients with genotype 2 or 3 and those with a viral load of less than 600,000 IU/mL have been found to achieve higher rates of sustained virologic response.<sup>15</sup> A sustained virologic response is defined as the absence of HCV RNA 24 weeks after stopping treatment and is now considered to be the best predictor of long-term treatment response. A sustained virologic response is generally regarded as a "virologic cure."

### HCV GENOTYPE AFFECTS SUCCESS AND DURATION OF TREATMENT

HCV has at least six major genotypes.<sup>1,3–6</sup> Several genotypes are subclassified as "a" or "b" (ie, genotype 1a or 1b); however, these distinctions are of little clinical use.

In the laboratory, HCV genotypes are identified by restriction fragment length polymorphism, by direct sequence analysis, or by

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reverse hybridization. Once the HCV genotype has been identified, there is no need to repeat the test.

Different genotypes are more common in some areas of the world than in others. Genotype 1 is the one most common in the United States (accounting for 70% to 75% of cases), followed by genotypes 2 and 3 (25%–30%). Genotype 4 is most common in Egypt and the Arabian peninsula.

HCV genotyping is important because it can help predict the likelihood of a response to treatment and in planning the dose and duration of therapy.<sup>77</sup> For example, treatment with pegylated interferon plus ribavirin is predicted to work approximately 50% of the time for people with genotype 1, but 80% to 90% of the time for people with genotypes 2 or 3.<sup>15–17,78</sup> Additionally, patients with genotype 1 need 12 months of therapy to achieve maximum benefit, whereas those with genotypes 2 and 3 require treatment for only 6 months to achieve maximum benefit.

Response rates to combination therapy for chronic HCV infection in a clinical setting and derivation of probability tables for individual patient management. J Viral Hepat 2008; 15:271–278.

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