Gastric intestinal metaplasia (GIM), a common histologic finding, is associated with increased risk of gastric cancer, and GIM associated with *Helicobacter pylori* infection is classified as an environmental metaplastic atrophic gastritis. Patients may be asymptomatic or present with various dyspeptic symptoms. Autoimmune metaplastic atrophic gastritis is a less common but important cause of chronic gastritis. The Correa cascade describes the evolution of precancerous mucosal changes that lead to development of GIM, with differentiation of 2 histologic types of GIM (complete and incomplete) and the consequences of each type. The risk of progression to malignancy is higher with incomplete GIM. It is also higher for those who immigrate from regions with a high incidence of *H pylori* infection to areas where the incidence is low. Guidelines regarding endoscopic management of GIM vary by geographic region.

**KEY POINTS**

Factors in the complex chain of events leading to malignant transformation include genetic predisposition, the anatomic extension of the metaplasia, and histologic differentiation.

Environmental risk factor control such as *H pylori* eradication, smoking cessation, and moderation in alcohol intake may halt the progression of atrophic gastritis to GIM.

Careful risk stratification is key: Patients at high risk should undergo endoscopic surveillance.

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Figure 1. Endoscopic appearance of gastric intestinal metaplasia (GIM). (A) White-light endoscopy reveals macroscopic GIM, with an irregular, even surface. The arrow indicates an elongated, groove-type pit pattern. (B) Enhanced narrow-band imaging of the same surface shows multiple pale, elevated patches.

### TABLE 1

<table>
<thead>
<tr>
<th></th>
<th>Autoimmune-mediated metaphlastic atrophic gastritis</th>
<th><em>Helicobacter pylori</em>-mediated metaphlastic atrophic gastritis</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Location</strong></td>
<td>Primary involvement of the gastric body and fundus</td>
<td>All gastric epithelium including antrum</td>
</tr>
<tr>
<td><strong>Acid production</strong></td>
<td>Lost entirely</td>
<td>Decreased</td>
</tr>
<tr>
<td><strong>Fasting gastrin level</strong></td>
<td>Markedly elevated</td>
<td>Variable</td>
</tr>
<tr>
<td><strong>Antibodies</strong></td>
<td>Antibodies to intrinsic factor and parietal cells</td>
<td>Antibodies to <em>H pylori</em></td>
</tr>
<tr>
<td><strong>Vitamin B₁₂ deficiency</strong></td>
<td>Present, often severe</td>
<td>May be present, usually mild</td>
</tr>
<tr>
<td><strong>Association</strong></td>
<td>Other autoimmune disorders</td>
<td>Peptic ulcer disease, adenocarcinoma, &quot;MALToma&quot;</td>
</tr>
</tbody>
</table>

MALToma = mucosa-associated lymphoid tissue lymphoma or MALT lymphoma

Based on information in reference 15.

progression risk. The overall risk of progression to esophageal adenocarcinoma in Barrett esophagus is 0.22% per year. The presence of low-grade dysplasia increases the annual cancer risk to 0.5% per year, and high-grade dysplasia increases the risk to 5% to 8% per year. Although the risk of cancer progression with GIM and Barrett esophagus is similar in the United States, endoscopic surveillance only improved patient-important outcomes in Barrett esophagus, likely because of the lower prevalence of GIM compared with Barrett esophagus.

### CHRONIC GASTRITIS AND GIM

Regardless of its cause, chronic gastric inflammation may lead to atrophic gastritis characterized by mucosal thinning and replacement of gastric glandular cells by intestinal epithelium (ie, GIM).

Environmental metaphlastic atrophic gastritis

*Helicobacter pylori* infection remains the leading cause of chronic gastritis, with earlier studies suggesting that it is responsible for more than 90% of cases.
GIM as a result of *H. pylori* infection is classified as an environmental metaplastic atrophic gastritis (EMAG). *H. pylori* is more prevalent than previously thought, based on estimates that 50% of the world population has been infected in their lifetime, and the overall prevalence in the United States is 36%. If not eradicated, *H. pylori* infection can progress to atrophic gastritis with damage to the gastric glands. Notably, the virulence of specific *H. pylori* strains can play a critical role in infection outcomes. Strains that express the cytotoxin-associated gene *CagA* or the vacuolating cytotoxin *VacA* *s1m1* genotype are associated with an increased risk of precancerous lesions and progression to adenocarcinoma.

Chronic use of proton pump inhibitors (PPIs) has not been shown to prevent or modify histologic changes of GIM. In fact, chronic PPI use often results in decreased *H. pylori* densities and proximal migration of the bacteria from the antrum to the body of the stomach, factors that complicate its diagnosis and timely eradication. Unmonitored long-term use of PPIs should be avoided.

Other possible causes of EMAG include habits such as high salt intake, cigarette smoking, and alcohol use.

Clinically, patients with EMAG may be asymptomatic or present with dyspeptic symptoms with variable severity. Autoantibodies to parietal cells and intrinsic factor are lacking, and levels of fasting gastrin tend to be low. In addition to evaluation for *H. pylori* and its timely eradication, EMAG patients should be screened for coexisting conditions such as vitamin B₁₂ and iron deficiency and treated appropriately.

**Autoimmune metaplastic atrophic gastritis**

A less common but important cause of chronic gastritis is autoimmune metaplastic atrophic gastritis (AMAG). Affecting 0.15% of the adult population, AMAG primarily involves the gastric body and fundus while sparing the antrum. Most patients are asymptomatic, but some may present with manifestations of vitamin B₁₂ deficiency or iron-deficiency anemia. In contrast to laboratory findings for EMAG, supportive laboratory findings with AMAG include positive antibodies to intrinsic factor (more specific) and parietal cells (more sensitive), fasting hypergastrinemia, and decreased serum pepsinogen I/II ratio. Screening should be considered for concomitant autoimmune conditions such as type 1 diabetes mellitus and autoimmune thyroid disease.

**Table 1** compares the features associated with EMAG and AMAG.
GASTRIC INTESTINAL METAPLASIA

The Correa cascade describes the progression from precancerous histologic changes in the gastric mucosa to the development of GIM and its consequences, including adenocarcinoma (Figure 2). The process begins with development of nonatrophic gastritis and progresses to multifocal atrophic gastritis followed by GIM.

GIM can present 2 histologic types:
- The complete and fully intestinalized pattern has markers for intestinal mucin and absence of gastric mucin.
- The incomplete gastric and intestinal mixed glands pattern has both gastric mucin and intestinal mucin markers.16

Complete GIM may progress to incomplete GIM if conditions leading to severe inflammation are present (eg, advanced atrophy or hypochlorhydria) before identifiable dysplastic changes.17 Subsequently, the tissue progresses to low-grade dysplasia, followed by high-grade dysplasia, and finally invasive adenocarcinoma.18

Differentiation of the 2 types of GIM is important. Incomplete GIM has been associated with an increased risk of cancer progression, and some experts consider it a mild degree of dysplasia.19,20

The risk of developing gastric cancer may be higher in patients with histologic evidence of incomplete and extensive GIM (ie, involvement of the antrum and corpus) than in those with complete and limited GIM.1,21,22 Some studies suggest that the topographic distribution of intestinal metaplasia may affect the risk of cancer progression. In Cassaro et al’s23 cohort study of 135 Colombian patients, a GIM distribution involving the lesser curvature of the stomach from the cardia to the pylorus was associated with higher cancer risk (odds ratio 5.7, 95% confidence interval 1.3–26) compared with “antrum-predominant” or “focal” patterns.23

The incidence of gastric cancer exhibits significant geographic variation worldwide due to potential environmental exposure factors and genetic predisposition. The reported rates are highest in Eastern Asia, Eastern Europe, and South America, and lowest in North America.24 People who immigrate from a region of high incidence to a region of low incidence have an increased risk of gastric cancer.25 Table 2 summarizes risk factors for malignancy.1,26

### TABLE 3

Variations in society recommendations for the management of gastric intestinal metaplasia

<table>
<thead>
<tr>
<th>Geographic location</th>
<th>Society recommendations</th>
</tr>
</thead>
<tbody>
<tr>
<td>East Asia</td>
<td>Endoscopic or radiographic screening of all men and women at age 50 or older29</td>
</tr>
<tr>
<td></td>
<td>In patients with gastric intestinal metaplasia and high-risk features, endoscopy recommended in 1 to 3 years</td>
</tr>
<tr>
<td>Europe</td>
<td>Patients with extensive gastric intestinal metaplasia should undergo endoscopic surveillance every 3 years</td>
</tr>
<tr>
<td></td>
<td>Consider endoscopic surveillance in patients with gastric intestinal metaplasia limited to the corpus or antrum of the stomach but with a family history of gastric cancer, persistent Helicobacter pylori infection, incomplete gastric intestinal metaplasia, or autoimmune gastritis30</td>
</tr>
<tr>
<td>United States</td>
<td>AGA recommends against routine endoscopic surveillance after gastric intestinal metaplasia is detected in the general population, but if H pylori is detected, treatment is encouraged</td>
</tr>
<tr>
<td></td>
<td>Patients with gastric intestinal metaplasia and risk factors associated with progression could be considered for endoscopic surveillance every 3 to 5 years if the patient favors surveillance (which has an unclear impact on mortality risk) vs endoscopic evaluation, which has a risk of complications31</td>
</tr>
<tr>
<td></td>
<td>ASGE recommends endoscopic surveillance exclusively in patients with risk factors, but not in the general cohort of patients in whom gastric intestinal metaplasia is detected32</td>
</tr>
</tbody>
</table>

AGA = American Gastroenterological Association; ASGE = American Society for Gastrointestinal Endoscopy

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**Correa Cascade: Differentiating Complete and Incomplete GIM**

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**Risk Factors for Progression to Malignancy**
ENDOSCOPIC MANAGEMENT

The role for endoscopy in GIM is limited to detection and surveillance, as no other methods are currently available for this. Specific recommendations for endoscopy are discussed in the various guidelines below.

NONENDOSCOPIC MANAGEMENT

GIM management should emphasize risk-factor modification, including smoking cessation and moderation in alcohol intake. In patients with \textit{H pylori}-induced gastritis, early \textit{H pylori} detection and eradication are crucial to halt progression to gastric cancer. In contrast, the effects of \textit{H pylori} eradication once GIM occurs are undetermined. GIM changes may be irreversible, and the impact of \textit{H pylori} eradication on cancer progression once GIM is established may be minimal.27

Observational studies have reported partial GIM reversal and decreased progression to stomach cancer with use of aspirin and nonsteroidal anti-inflammatory drugs such as celecoxib.28 More evidence is needed to support their use.

GLOBAL DIFFERENCES IN GUIDELINE RECOMMENDATIONS

The optimal follow-up of patients with isolated glands of GIM remains controversial, with significant differences in guidelines in the Eastern and Western regions of the world (Table 3).29-32

Figure 3. An algorithmic approach to the management of gastric intestinal metaplasia (GIM).
GASTRIC INTESTINAL METAPLASIA

Recommendations for Eastern regions
In countries such as Japan, where the incidence of gastric cancer is high, national screening programs recommend mass endoscopic or radiographic screening of all men and women at age 50 or older. For patients found to have GIM without malignancy on initial screening, surveillance endoscopy in 1 to 3 years is recommended if they have GIM with high-risk features such as incomplete GIM, extensive GIM, family history of gastric cancer, smoking, or excessive alcohol use.

Recommendations for Western regions
The British Society of Gastroenterology guidelines identify patients with GIM as having increased risk for gastric malignancy and recommend endoscopic surveillance every 3 years if there is extensive GIM (ie, affecting the antrum and corpus), antral GIM with risk factors such as H pylori, or a family history of gastric cancer.

A 2019 consensus guideline by 4 European organizations agreed with the British Society of Gastroenterology. The guideline recommends that patients with GIM who are considered at high risk, including those with histologically proven GIM of the corpus and antrum, undergo endoscopic surveillance every 3 years. The guideline advises consideration of surveillance if GIM is present only in the corpus or antrum but the patient has a family history of gastric cancer, persistent H pylori, incomplete GIM, or autoimmune gastritis.

US recommendations
Two US societies have published guidelines addressing the management of GIM.

The American Gastroenterological Association guidelines recommend against routine endoscopic surveillance after GIM is detected in the general population, but if H pylori is detected, treatment is encouraged. Patients with GIM and risk factors associated with progression can be considered for endoscopic surveillance every 3 to 5 years if the patient favors surveillance, which has an unclear impact on mortality risk, vs endoscopic evaluation, which has potential complications.

The guidelines subcategorized risk factors associated with progression of gastric cancer as follows:

- Highest risk: incomplete GIM, extensive GIM, or family history of gastric cancer
- Overall increased risk: certain racial or ethnic minorities immigrating from high-incidence regions

The American Society for Gastrointestinal Endoscopy recommendations are similar to those of European groups. They advise endoscopic surveillance exclusively for patients with risk factors, but not for the general cohort of patients in whom GIM is detected.

AN ALGORITHMIC APPROACH TO DIAGNOSIS AND MANAGEMENT

Figure 3 suggests an approach to managing patients who have GIM. The updated Sydney protocol includes the collection of 5 nontargeted biopsy specimens: 2 from the antrum (at the lesser and greater curvature), 2 from the corpus (at the lesser and greater curvature), and 1 from the incisura. It is recommended that these biopsy specimens be placed in separate jars.

Careful inspection should be carried out with high-definition white-light endoscopy rather than standard-definition endoscopy. Adequate air insufflation, use of mucolytic and defoaming agents (for improved visibility), appropriate withdrawal times, and photodocumentation are key for a quality endoscopic examination. Additionally, use of narrow-band imaging should be encouraged because it has been shown to improve the detection of GIM. It also allows for more targeted biopsies for GIM.

DISCLOSURES

The authors report no relevant financial relationships which, in the context of their contributions, could be perceived as a potential conflict of interest.

REFERENCES


